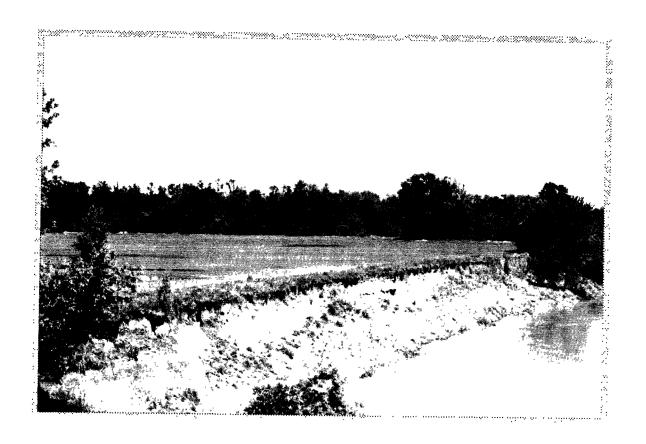
Open-File Report 95-156

COLLECTION OF SHORT PAPERS ON THE BEAVER CREEK WATERSHED STUDY IN WEST TENNESSEE, 1989-94



Prepared by the U.S. GEOLOGICAL SURVEY





Organizations involved in assessment studies in the Beaver Creek area:

U.S. Department of Agriculture, Natural Resources Conservation Service (formerly known as Soil Conservation Service)

U.S. Department of the Interior, U.S. Geological Survey

Tennessee Department of Agriculture

Tennessee Department of Environment and Conservation

Shelby County Conservation District

University of Tennessee, Agricultural Extension Service

Clemson University

The University of Memphis

Cover photograph. Loosahatchie River looking upstream at Highway 205 in the Beaver Creek watershed.

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Compiled by W. Harry Doyle, Jr., and Eva G. Baker

U.S. GEOLOGICAL SURVEY

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Memphis, Tennessee 1995

U.S. DEPARTMENT OF THE INTERIOR BRUCE BABBITT, Secretary

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PREFACE

In 1989, the U.S. Geological Survey began a scientific investigation to evaluate the effect of agricultural activities on water quality and the effectiveness of agricultural best management practices in the Beaver Creek watershed, West Tennessee. The project is being conducted jointly with other Federal, State, County agencies, the farming community, and academic institutions, in support of the U.S. Department of Agriculture's Hydrologic Unit Area program.

The Beaver Creek project has evolved into a long-term watershed assessment and monitoring program. In 1991, a grant was received to develop and evaluate sampling strategies for higher order streams. During the summer of 1992, a reconnaissance of water-quality conditions for the shallow aquifers in Shelby, Tipton, Fayette, and Haywood Counties was conducted and included 89 domestic wells in the Beaver Creek watershed. Results from this effort lead to the development of a 1-year program to evaluate cause-and-effect relations that can explain the observed water-quality conditions for the shallow aquifers in the watershed. In 1992 the USGS, in cooperation with the Soil Conservation Service and the Shelby County Soil Conservation District, began an evaluation of in-stream processes and in-stream resource-management systems. In 1993, a biomonitoring program was established in the watershed.

This collection of articles and abstracts was originally published in the American Water Resources Association National Symposium on Water Quality Proceedings for the National conference held in Chicago in 1994. These articles address the optimimum sampling strategy for obtaining water-quality data in nonpoint-source pollution studies; provide an understanding of the fate and transport of agrichemicals in the surface runoff and the soil profile; demonstrate that the most significant water-quality problem in the watershed is the transport of sediment; support qualitative and quantitative statements of the effects agriculture has on water quality in the Beaver Creek watershed. The purpose of this report is to present the current (1994) results of these investigations in one document related to the Beaver Creek watershed.

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ASSESSMENT OF AGRICULTURAL NONPOINT-SOURCE POLLUTION AND BEST MANAGEMENT PRACTICES FOR THE BEAVER CREEK WATERSHED, WEST TENNESSEE: AN OVERVIEW

Angel Roman-Mas¹, Robert W. Stogner, Sr.¹, W. Harry Doyle Jr.¹, and Stephen J. Klaine²

ABSTRACT: In 1989, the U.S. Geological Survey began a long-term research project to evaluate the effect of agricultural activities on water quality and the effectiveness of agricultural best management practices in the Beaver Creek watershed, West Tennessee. The project is being conducted jointly with other Federal, State, and county agencies, the farming community, and academic institutions, in support of the U.S. Department of Agriculture Hydrologic Unit Area program. This paper presents a historical account of the project, a general description of the study area and the methods used for data collection and analyses, and a synopsis of the most significant project findings.

KEY TERMS: Agriculture, nonpoint-source pollution; best management practices; water quality; ground water; surface water; soils.

INTRODUCTION

In 1989, efforts by the farming community in Tennessee facilitated a cooperative agreement between the U.S. Geological Survey (USGS) and the Tennessee Department of Agriculture (TDA) to evaluate agricultural nonpoint-source pollution and best management practices (BMP's) in the Beaver Creek watershed, West Tennessee. An assessment conducted by the Tennessee Department of Environment and Conservation (TDEC) (formerly the Tennessee Department of Health and Environment, 1988) indicated that agricultural activities were the principal cause for water-quality degradation in this watershed. This paper presents a historical account of the project, a general description of the study area and the methods used for data collection and analyses, and a synopsis of the most significant project findings.

For this study, a multidisciplinary, multiagency team of scientists from the USGS, Clemson University, University of Memphis, and the University of Tennessee Agricultural Extension Service (UTAES) was assembled. The team selected four small basins (less than 50 hectares each) to conduct deterministic (cause-and-effect relation) and mechanistic (processes and factors) evaluations of agricultural nonpoint-source pollution and BMP's. A major component of this study was the evaluation of existing sampling methods for runoff and soils and the development of new methods as needed.

In 1990, the Beaver Creek watershed was selected for the U.S. Department of Agriculture (USDA) Hydrologic Unit Area (HUA) program. The HUA program provides technical, financial, and educational assistance to farmers for the implementation of BMP's. The following BMP's were identified: (1) conservation tillage systems, (2) terrace systems, (3) conservation reserve program, (4) contour stripcropping, (5) diversions, (6) critical area treatment, (7) grass waterways, and (8) water and sediment control basins.

¹Hydrologist, U.S. Geological Survey, 7777 Walnut Grove Rd., Suite LLB2, Memphis, Tennessee 38120.

²Associate Professor, Department of Environmental Toxicology and The Institute of Wildlife and Environmental Toxicology, Clemson University, Pendleton, South Carolina, 29670-0709.

The Beaver Creek project has evolved into a long-term watershed research and monitoring program. In 1991, a grant was received from the Water Environment Federation to develop and evaluate sampling strategies for higher order streams. During the summer of 1992, a reconnaissance of water-quality conditions for the shallow aquifers in Shelby, Tipton, Fayette, and Haywood Counties was conducted (Fielder and others, 1994). The reconnaissance, which was undertaken in cooperation with the TDA and the UTAES, included 89 domestic wells in the Beaver Creek watershed. Results from this effort lead to the development of a 1-year program to evaluate cause-and-effect relations that can explain the observed water-quality conditions for the shallow aquifers in the watershed. The evaluation is being funded by the TDA, the TDEC, and the Shelby County Conservation District (SCCD). In 1992 the USGS, in cooperation with the Soil Conservation Service and the SCCD, began an evaluation of in-stream processes and in-stream resource-management systems. In 1993, a biomonitoring program was established in the watershed.

Current project objectives include the: (1) evaluation of the extent that agricultural activities affect water quality and threaten environmental integrity in the Beaver Creek watershed, (2) evaluation of in-field and instream BMP's, (3) development and evaluation of chemical and biological sampling strategies, (4) identification and quantification of processes and factors that control the degradation and transport of agricultural pollutants, and (5) evaluation of mathematical-mechanistic models. Results from the Beaver Creek watershed project can assist resource management agencies in developing strategies for use in their conservation programs.

DESCRIPTION OF STUDY AREA

The Beaver Creek watershed, located in Shelby, Tipton, Haywood, and Fayette Counties in southwest Tennessee (see Figure 1), encompasses about 40,000 hectares (ha). The watershed lies within the Southern Mississippi Valley Silty Uplands major land resource area (U.S. Department of Agriculture, Soil Conservation Service, 1988). Two major physiographic regions, upland and bottomland, can be distinguished in the watershed. Agriculture is vital to the local economy. Cropland covers about 70 percent of the watershed; cotton, soybeans, and corn are the principal crops. Forests and range land occupy 10 and 20 percent of the watershed, respectively. Forest stands are primarily pole and small saw timber, reflecting extensive harvesting 40 to 60 years ago. All major tributaries in the Beaver Creek watershed were channelized in the early to mid-1900's to facilitate drainage.

The mean annual rainfall is about 130 centimeters (cm); about 25 cm fall during the 180-day growing season of April through September. The wettest month is April, with a mean rainfall of 14 cm, and the driest month is October, with a mean of 6 cm. Mean monthly temperatures range from 1 °C (January) to 27 °C (July) and the mean annual temperature is 16 °C.

Geologically, the watershed is in the Mississippi embayment of the Gulf Coastal Plain physiographic province. Alluvium and fluvial deposits of Quaternary and Tertiary(?) age make up the shallow aquifer in the watershed (Parks, 1990; Parks and Carmichael, 1990a, 1990b). Alluvium, consisting of sand, gravel, silt, and clay, is present beneath the bottomland areas; fluvial deposits of sand, gravel, minor clay, and ferruginous sandstone are present beneath the loess in the uplands and valley slope areas (Parks, 1990; Parks and Carmichael, 1990a, 1990b). Locally, the alluvium and fluvial deposits are underlain by the Cockfield Formation, Cook Mountain Formation, or the Memphis Sand of Tertiary age. The Cockfield Formation and the Memphis Sand consist of sand, silt, clay, and lignite and make up the Cockfield and Memphis aquifers (Parks, 1990; Parks and Carmichael, 1990a, 1990b). The Cook Mountain Formation, which consists primarily of clays, makes up the confining unit overlaying the Memphis aquifer.

Soils in the watershed consist of well to poorly drained silt loam (U.S. Department of Agriculture, Soil Conservation Service, 1988). Three principal soil associations are present: Memphis-Loring, Granada-Loring-Memphis, and Falaya-Waverly-Collins. The Falaya-Waverly-Collins soil association occurs in the bottomland areas whereas the other two soil associations are typically in upland areas. The soils are acidic with pH ranging from 5.1 to 5.5 units.

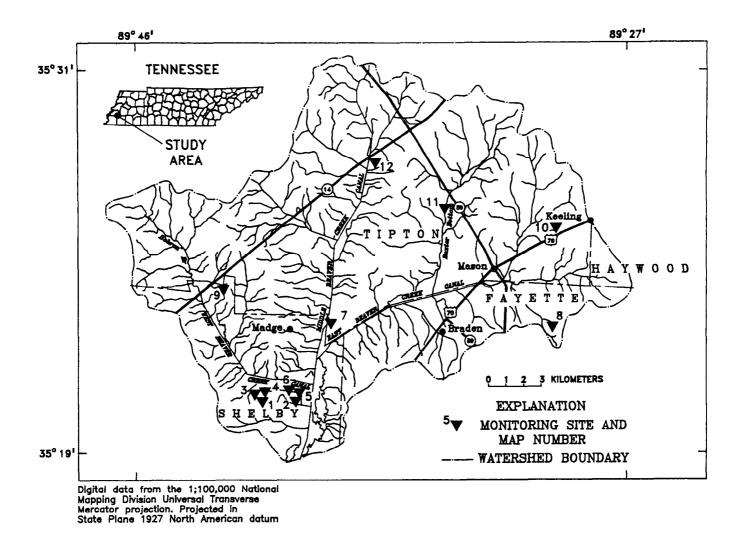


Figure 1. Location of the Beaver Creek watershed study area and surface-water monitoring sites.

RESEARCH AND MONITORING ACTIVITIES

For the Beaver Creek watershed project, a watershed-based risk assessment approach was adopted to design the research and monitoring activities. The following steps were taken as part of this approach:

- (1) Identification of the natural resources and their specific functions: For the Beaver Creek project, this task was conducted by the SCS in support of the HUA program. Soils and water quality were the identified resources. Specific functions of these resources are production agriculture, fish and wildlife, aquatic recreation activities, irrigation, and livestock.
- (2) Identification of stressors: This step refers to anthropogenic activities that can degrade the resources or impair specific functions identified in step 1. Row-crop production (cotton, soybean, corn), cattle (range land), and channelization were identified as potential stressors.
- (3) Assessment of physiographic variations: The watershed-based risk assessment approach requires identification of the range of physiographic settings that can produce different resource-stressor or function-stressor responses. For instance, in the Beaver Creek watershed, cotton and soybeans are cultivated in upland and bottomland areas. Therefore, the evaluation of cotton and soybean crops as a potential stressor must include experimental fields located in both these areas.

- (4) Evaluation of BMP's: The evaluation of BMP's adds another dimension to the monitoring program. Paired-basins analysis, which is the preferred technique to evaluate BMP's, required two monitoring sites for every resource-stressor response identified in a given physiographic setting. The upstream-downstream approach also requires two monitoring sites for every resource-stressor response to be studied. In this approach, data from a site downstream from the area where the BMP's have been implemented are compared to data collected at a site upstream from the BMP area. Trend analysis is the third and most common approach for evaluating BMP's. In this approach, pre- and post-BMP's implementation data are collected and compared. This approach typically includes one monitoring site at the downstream end of the area where the BMP's are implemented. In some instances, monitoring sites upstream and downstream of the BMP implementation area are needed to isolate the area's contribution to water-quality conditions.
- (5) Selection of monitoring sites: The number of sites to be monitored is based on the number of resource-stressors identified in the watershed and the BMP's to be evaluated. In the Beaver Creek watershed, row-crop agriculture and channelization were determined to be the most important stressors to soils and water resources in the watershed, and non-tillage and riparian zone restoration were the most important BMP's. The scale of the monitoring sites is dictated by the type of investigation being conducted. Whereas occurrence and distribution studies can be conducted in relatively large-scale systems, mechanistic and deterministic evaluations typically require small-scale systems. Both types of evaluation are required to fulfill the Beaver Creek project objectives.
- (6) Data collection: Any biases imposed by the data-collection methods must be recognized and quantified. Sensitivity analyses, in which the magnitude of response of a parameter is evaluated relative to changes in other parameters, are necessary to develop, evaluate, and modify data-collection methods.
- (7) Modeling: Modeling efforts should provide resource managers with tools (1) to simulate field- and watershed-level processes related to specific stressor-resource response and (2) to extend information gained in bench-mark studies to other areas. Models can range in approach from synoptic, which provide a broad overview of the resource-stressor response with minimum data, to mathematical-mechanistic, which attempt a full differentiation of the processes and factors controlling the resource-stressor response.

Surface Water

Twelve surface-water monitoring sites have been established in the watershed (see Figure 1). Climatic, hydrologic, chemical, biological, and geomorphological (channel evolution) data collected at these sites are being used in paired-basin analyses, upstream-downstream comparisons, and trend analyses (see Table 1).

Streamflow Stations and Measurements

Each surface-water monitoring site includes a stream-gaging station to measure discharge. The stations were constructed and are being operated according to standard USGS procedures. Drainage boundaries for each station were determined and the drainage areas computed. Topographic maps for each basin were developed from 7.5-minute quadrangle topographic maps or from field surveys. A detailed inventory of farm activities is being maintained.

Each station was outfitted with a pressure sensor or a stilling well and an automated recorder programmed to record stream elevation (stage) at variable time intervals according to the rate of change in stage. Continuous discharge records were developed from the stage data following standard USGS methods (Kennedy, 1983). At selected monitoring sites (see Figure 1, sites 1, 8, 9, and 10) automated rain gages were installed.

Unit hydrograph analysis and water budgets are being used to evaluate the hydrologic response of the basins. It is reasonable to expect that changes in the hydrologic response of the basins would be observed if the implementation of BMP's are effective. Increases in infiltration rates and evapotranspiration and associated decreases in runoff to rainfall ratio, basin response time, and peak discharge are expected to result from the implementation of BMP's.

Table 1. Location, data collection, physiographic and agricultural information for surface-water monitoring stations in the Beaver Creek watershed, Tennessee

[hyd, hydrological; cli, climatic; chm, chemical; bio, biological; geo, gemorphological; *, estimated from topographic maps]

Map Number	Data Being Collected	Type of Evaluation Being Conducted	Drainage Areas (hectares)	Area Under Cultivation (percent)	Crop(s)	Physiography
1-2	hyd, eli, chm, bio, and geo	Up-stream/down-stream, in-stream processes, and resources management Sampling protocols	171-258	95-95	Cotton soybean	Bottomland
3-4	hyd, cli, chm, and geo	Paired-basins, conventionally tilled versus non-tilled cotton	6.88-6.93	100-100	Cotton	Bottomland
5-6	hyd, cli, chm, and bio	Up-stream/down- stream, constructed wetland	0.4 (wetland)- 28 (cropland)	0- 100	Soybean com	Bottomland
7	hyd, cli, chm, and geo	 Trend analyses Sampling protocols 	12,000*	65	Cotton	Upland and bottomland
8	hyd, cli, chm, and geo	 Trend analyses Sampling protocols 	44	100	Soybeans	Upland
9	hyd, cli, chm, and geo	Trend analyses Sampling protocols	26	80	Soybeans	Upland
10	hyd, cli, chm, and geo	 Trend analyses Sampling protocols 	11	100	Cotton	Upland
11-12	hyd, cli, chm, bio, and geo	 Paired basin, channelized versus non-channelized Sampling protocols 	1,215-1,135°	70 -70	Cotton soybeans	Upland

Water Quality

Surface-water samples for the determination of nitrogen and phosphorous species, organic carbon, suspended sediment, and selected pesticides and their metabolites are being collected during stormflow and base-flow conditions. Two automated samplers were installed at each station to aid in the sample collection. Approximately 20 rainfall-runoff events are monitored annually. During stormflow, water samples are collected at an interval equal to 5 percent of the total stormflow duration. During base-flow, water samples are collected every 15 minutes for the first 3 hours after stormflow and every hour thereafter until the end of the event or the beginning of a new event.

Annual and seasonal concentration-frequency curves (percent of the time that a particular concentration is exceeded during a given period of time) are being developed for each basin. Loads for annual, seasonal, and individual rainfall-runoff events are being computed. Statistical analyses will be used to determine if the variance in concentration-frequency curves and loads could be attributed to the implementation of BMP's. In addition, changes in chemical speciation and the phase (suspended or dissolved) in which the constituents occur are being evaluated.

Coarse particulate organic matter (CPOM) in surface water is being sampled by diverting approximately 8 percent of the total flow into a collection net. Trapped CPOM is collected, placed in a labeled plastic bag, and transported to the laboratory after each storm event. CPOM samples are being analyzed for particle size and for nitrogen, phosphorous, and pesticide content. Differences in the physical, chemical, and biological characteristics of the CPOM that determine its suitability as a food source for organisms downstream are also being evaluated.

Mathematical models that can simulate runoff and constituent transport at the field and watershed levels [for example, AGNPS (Young and others, 1991) and HSPF (Donigian, 1991)] will be calibrated and applied to the basin being monitored. A detailed evaluation of the mathematical algorithms used in these models will be conducted and modified as needed. Only those models that attempt a full differentiation of the processes and factors controlling the degradation and transport of agricultural chemicals are being considered.

Benthic macroinvertebrate data are being collected at selected sites in the Beaver Creek watershed (see Figure 1 and Table 1). The primary goal of the biomonitoring program is to evaluate and develop, as needed, biological sampling methods for streams in the watershed. Various methods are currently being studied. In addition, structural and functional indices are being used to evaluate water-quality conditions and environmental integrity.

In-Stream Processes

Modifications to the drainage network to improve drainage and control floods have altered the hydrologic regime of streams and marginal ecosystems (such as wetlands) in the Beaver Creek watershed. The preservation of biological and non-biological functions (for example, habitat and water quality, respectively) performed by streams and marginal ecosystems is central to the development of environmentally sound agriculture in the watershed. Quantitative understanding of these functions and the extent to which they have been impaired by drainage modifications is needed before in-stream resource-management systems can be developed and implemented.

Although little information is available on the natural drainage system in the bottomlands of the Beaver Creek watershed, the system was likely dominated by forested riverine wetlands. Flooding conditions of the bottomlands hampered their exploitation for timber and agriculture and lead local landowners to seek engineering solutions to drainage problems (Ashley, 1910). Channelization has been the most common engineering practice used in the watershed.

The Upper Middle Beaver Creek Canal subwatershed (see Figure 1, monitoring site 12) represented a typical example of a channelized condition. In contrast, the Baxter Bottom subwatershed at monitoring site 11 (see Figure 1) has not been channelized. A 250-ha forested riverine wetland is part of the Baxter Bottom subwatershed drainage network. This condition likely resembles natural drainage conditions for bottomlands in the Beaver Creek watershed. The two subwatersheds are of comparable size, slope, and land use. In 1993, a paired-basin study was started to document differences in water quality between the two subwatersheds and to identify and quantify biological, geochemical, and geomorphological processes responsible for the observed differences in water quality.

Constructed Wetlands

Wetlands can immobilize and metabolize chemical constituents through a complex network of physical, chemical, and biological processes and thereby improve water quality at their downstream ends. Constructed wetlands have been proposed as an off-site BMP for the treatment of cropland runoff because in many instances the transport of agricultural chemicals in runoff cannot be controlled in the field. Constructed wetlands have been successfully used to treat domestic and animal wastewater (Watson and others, 1989).

The design parameters used in constructed wetlands for the treatment of wastewater are typically determined by reaction kinetics (Kadlec, 1989; Watson and others, 1989). These parameters are unsuitable for designing wetlands to be used for treating cropland runoff because of the extreme temporal variations in flow and constituent concentrations, the chemical speciation of the constituents, and the unknown reaction mechanisms and rates for these chemical species in wetland environments.

In 1993, a 0.4-ha single-cell herbaceous wetland was constructed in the Beaver Creek watershed. Its purposes are to (1) evaluate the effectiveness of the wetland in assimilating or modifying the nutrient and pesticide content in cropland runoff; (2) identify and quantify the relevant processes responsible for the immobilization and metabolization of selected constituents within the wetland; and (3) develop design criteria for constructed wetlands to be used for treating cropland runoff.

Stream-gaging and water-quality monitoring stations were established upstream and downstream of the wetland (see Figure 1, monitoring sites 5 and 6). Water samples are being collected and analyzed for the determination of total and dissolved nitrogen and phosphorous species, organic carbon, selected pesticides and their metabolites, and suspended-sediment concentration. Changes with time in the concentration of these constituents, chemical speciation, and loads at the upstream and downstream sampling stations are being compared. Biological samples are being collected within the wetland for the determination of succession patterns in the benthic macroinvertebrate and the plant communities.

Soils

Non-tillage has been effectively used to reduce soil erosion. However, researchers have suggested that non-tillage can enhance chemical transport through the soil zone and increase the potential for ground-water contamination (Dick and others, 1989; Hall and others, 1989). Resource management agencies in the Beaver Creek watershed, which has soils that are highly susceptible to erosion, have recommended non-tillage as a BMP. The potential for ground-water contamination associated with the implementation of non-tillage in the watershed needs to be addressed before non-tillage is widely adopted.

Since 1990, a series of experiments have been conducted to (1) develop a sampling strategy that accurately characterizes the spatial and temporal distribution of agricultural chemicals in the soil profile, (2) evaluate the transport and degradation of agricultural chemicals in the soil profile, (3) compare the behavior of agricultural chemicals in conventionally tilled and non-tilled cotton fields, and (4) evaluate solute transport models such as CMLS (Nofziger and Hornsby, 1987), MOUSE (Steenhuis and others, 1987), PRZM (Carsel and others, 1985), GLEAMS (Leonard and others, 1987), and LEACHMP (Wagenet and Hutson, 1987).

Ground-Water Quality

As in many rural areas in the United States, residents in the Beaver Creek watershed rely on domestic wells for their primary source of water. Residents share a growing concern that agricultural activities could be affecting the quality of ground water in the watershed.

Water-quality, well-construction, and land-use data for 89 domestic wells (See Figure 2) completed within the shallow aquifers in the Beaver Creek watershed were used to statistically relate water-quality conditions to land-use activities. The data are part of a ground-water-quality reconnaissance conducted during the summer of 1992 for the shallow aquifers in Shelby, Tipton, Fayette, and Haywood Counties (Fielder and others, 1994). Non-parametric statistical methods were used to determine the extent to which land use affects the occurrence and distribution of nitrate within the water-table aquifer in the Beaver Creek watershed.

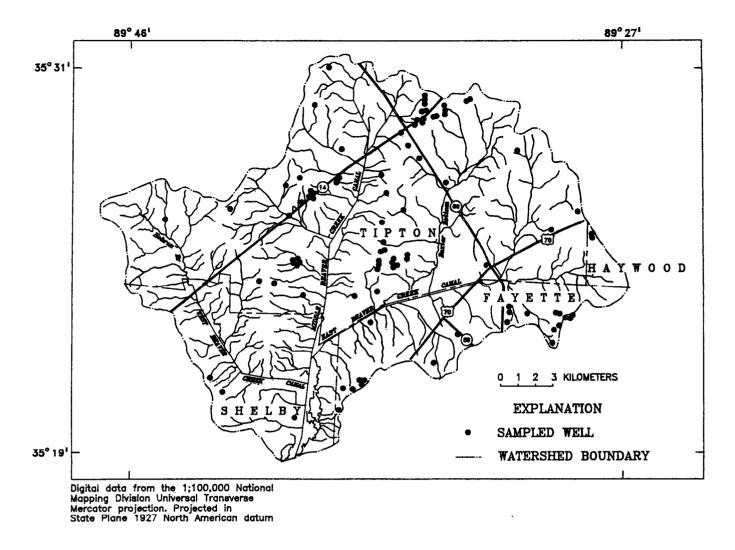


Figure 2. Location of sampled wells in the Beaver Creek watershed, 1992.

In 1993, a ground-water-quality network was established in the watershed. The network includes about 100 shallow wells that are sampled 5 times a year. Water samples are being analyzed for major constituents, nutrients, fecal bacteria, and selected pesticides. The water-quality data being collected will allow further investigation of cause-and-effect relations controlling the occurrence of pollutants in the shallow aquifers.

SIGNIFICANT FINDINGS

The following is a synopsis of the project's most significant findings.

- (1) An optimal sampling strategy for the temporal characterization of chemicals and suspended sediment in agricultural runoff includes frequent sampling during stormflow and less frequent sampling during recession or base flow. Sensitivity analyses showed that a sampling interval equal to 5 percent of the stormflow duration is required during stormflow. For recession or base flow, a calibrated regression can supplement a limited number of concentration measurements.
- (2) Soil-sampling transects perpendicular to the orientation of the rows provided adequate spatial representation to conduct statistical analyses and compute reliable field averages for the concentration of agricultural chemicals in soils.

- (3) Surface-catalyzed reactions appear to be an important mechanism for the degradation of aldicarb metabolites in the soil profile. The calculated half-lives of aldicarb metabolites were an order of magnitude shorter than those computed from laboratory experiments in which surface-catalyzed reactions were absent.
- (4) Horizontal transport within the soil profile was negligible for the conventionally tilled and the non-tilled fields.
- (5) Observed patterns in the concentration of aldicarb and its metabolites in runoff are consistent with the convective-diffusion limited transfer concept and inconsistent with the instantaneous-equilibrium concept used in lumped-parameter models.
- (6) Sediment transport resulting from soil erosion was identified as the major water-quality problem in the watershed. Computed suspended-sediment loads for these basins exceeded 30 metric tons per ha.
- (7) Crop residue at different stages of decomposition accounted for about 80 percent of the total nitrogen exported from crop fields. The chemical speciation and the temporal distribution of nitrogen exported from these basins could not be directly related to fertilizer application.
- (8) Background nitrate concentrations in water from the shallow aquifers have increased as a result of fertilizer application in the watershed. Significantly higher nitrate concentrations were measured in water from wells near septic tanks and confined animal facilities.
- (9) Community structure indices for benthic macroinvertebrates varied greatly depending on the sampling techniques used. Potential biases resulting from the sampling technique can be reduced by using multiple sampling techniques.

REFERENCES

- Ashley, G. H., 1910. Drainage problems in Tennessee. Tennessee State Geological Survey Bulletin 3-A, pp. 1-15.
- Carsel, R. F., L. A. Mulkey, M. N. Lorber and L. B. Baskin, 1985. The Pesticide Root Zone Model (PRZM): Procedure for Evaluating Pesticide Leaching Threats to Groundwater. Ecological Modelling 30:49-69.
- Dick, W. A., R. J. Roseberg, E. L. McCoy and W. M. Edwards, 1989. Surface Hydrological Response of Soils to Notillage. Soil Science Society of America Journal 53:1520-1526.
- Donigian, A. S., Jr., 1984. Application Guide for Hydrological Simulation Program--Fortran (HSPF). U.S. Environmental Protection Agency, Environmental Research Laboratory, Athens, GA, EPA/600/63/83/065.
- Fielder, A. M., A. Roman-Mas and M. W. Bennett, 1994. Reconnaissance of Ground-Water Quality at Selected Wells in the Beaver Creek Watershed, Shelby, Fayette, Tipton, and Haywood Counties, West Tennessee, July and August 1992. U.S. Geological Survey Open-File Report 93-366, 28 pp.
- Hall, J. K., M. R. Murray and N. L. Hartwig, 1989. Herbicide Leaching and Distribution in Tilled and Untilled Soil. Journal of Environmental Quality 218:439-445.
- Kadlec, R. H., 1989. Hydrologic Factors in Wetland Water Treatment, in Hammer, D. A., ed., Constructed Wetlands for Wastewater Treatment: Municipal, Industrial, and Agricultural. Lewis Publishers, Chelsea, MI, pp. 21-40.
- Kennedy, E. G., 1983. Computation of Continuous Records of Streamflow. U.S. Geological Survey Techniques of Water-Resources Investigations, Book 3, chapter A13, 53 pp.
- Leonard, R. A., W. G. Knisel and D. A. Still, 1987. GLEAMS: Ground-Water Loading Effects of Agricultural Management Systems. Transactions of the American Society of Agricultural Engineers 30:1403-1418.
- Nofziger, D. L. and A. G. Hornsby, 1987. Chemical Movement in Layered Soils: User's Manual. Florida Cooperative Extension Services, Institute of Food and Agricultural Sciences, University of Florida, Gainesville, FL, Circular 780, 44 pp.
- Parks, W. S., 1990. Hydrogeology and Preliminary Assessment of the Potential for Contamination of the Memphis Aquifer in the Memphis Area, Tennessee. U.S. Geological Survey Water-Resources Investigations Report 90-4092, 39 pp.
- Parks, W. S. and J. K. Carmichael, 1990a. Geology and Ground-Water Resources of the Cockfield Formation in Western Tennessee. U.S. Geological Survey Water-Resources Investigations Report 88-4181, 17 pp.
- Parks, W. S. and J. K. Carmichael, 1990b. Geology and Ground-Water Resources of the Memphis Sand in Western Tennessee. U.S. Geological Survey Water-Resources Investigations Report 88-4182, 30 pp.
- Steenhuis, T. S., S. Pacenka and K. S. Porter, 1987. MOUSE: A Management Model for Evaluating Groundwater Contamination from Diffuse Surface Sources Aided by Computer Graphics. Applied Agricultural Research 2:277-289.
- Tennessee Department of Health and Environment, 1988. Status of Water Quality in Tennessee. Tennessee Department of Health and Environment, Division of Water Management, Nashville, TN., 156 pp.

- U.S. Department of Agriculture, Soil Conservation Service, 1988. Drainage Basin Plan-Environmental Assessment, Beaver Creek Drainage Basin, Tennessee. [U.S.] Department of Agriculture, Soil Conservation Service, Nashville, TN, 74 pp.
- Wagenet, R. J. and J. L. Hutson, 1987. LEACHMP, Leaching Estimation and Chemistry Model. Center for Environmental Research, Cornell University, Ithaca, NY, 50 pp.
- Watson, J. T., S. C. Reed, R. H. Kadlec, R. L. Knight and A. E. Whitehouse, 1989. Performance Expectations and Loading Rates for Constructed Wetlands, in Hammer, D. A., ed., Constructed Wetlands for Wastewater Treatment: Municipal, Industrial, and Agricultural. Lewis Publishers, Chelsea, MI, pp. 319-350.
- Young, R. A., C. A. Onstad, D. D. Bosch and W.P. Anderson, 1991. Agricultural Non-Point Source Pollution Model, User's Guide, version 3.65. U.S. Department of Agriculture, Agriculture Research Services, Morris, Minnesota, 57 pp.

EVALUATION OF BIOMONITORING TECHNIQUES USED IN ASSESSING AGRICULTURAL NONPOINT SOURCE POLLUTION

Thomas D. Byl¹ and Angel Roman-Mas²

ABSTRACT: A biomonitoring program was established in the Beaver Creek Hydrologic Unit Area, West Tennessee in 1993, as part of an ongoing comprehensive water quality assessment program. A primary goal of the biomonitoring project was to evaluate and develop biomonitoring protocols appropriate for first- and second-order streams in the Beaver The biomonitoring techniques examined included Creek watershed. benthic community structure, colonization of multiplate samplers and plant debris packs, and decomposition of the plant material. Results varied greatly between biomonitoring techniques at the same sites. multiplate samplers generally had the lowest number of organisms and the smallest values for diversity. The plant debris packs and the benthic community structure techniques both provided high numbers for abundance and diversity, but under different conditions. The plant debris packs had a greater total number for abundance in the open streams than the shaded streams, whereas the benthic community structure-technique had higher numbers for abundance in the shaded streams than the open streams. These trends demonstrate a bias in the sampling techniques that should be considered when designing a biomonitoring project. Biases could be minimized by applying more than one biological collection technique to the same site.

KEY TERMS: Biomonitoring; channelized streams; macroinvertebrate; agriculture.

INTRODUCTION

Increased concern for water quality has resulted in the escalated use of biomonitoring to determine the impact of non-point pollution derived from agricultural activities, a major contributor of pollution in the USA (USEPA, 1989; USEPA, 1992). Biomonitoring is a technique that utilizes the natural biological community in the ecosystem to assess the impact of environmental stress. If a pollution problem exists, one expects to find a corresponding adverse effect on the aquatic biological community. The ecosystem can also be modified by human-induced channelization of streams, as has been done in many of the riverine systems in the southeast and southwest. The hydrologic and geomorphic modifications resulting from channelization can affect the biological community as much as pollution can (Cooper, 1993). Thus, aquatic biomonitoring techniques that were developed for study of natural settings must be validated and adapted before they are applied to these modified aquatic systems. Several biomonitoring techniques were evaluated to determine their effectiveness for assessing water quality in a modified stream of West Tennessee.

¹Research Biologist, U.S. Geological Survey, Water Resources Division, 810 Broadway, Suite 500, Nashville, TN 37203

²Research Hydrologist, U.S. Geological Survey, Water Resources Division, 7777 Walnut Grove, Suite LLB2, Memphis, TN 38120

The site selected for study was the Beaver Creek Watershed (BCW), located in the West Tennessee section of the Mississippi embayment. It is an area intensively farmed with row crops of cotton, soybean, winter wheat and corn. Soils in the BCW are predominately alluvial silt deposits in the bottomlands and loess deposits in the uplands. Recently, (1989-present) conservation practices have been initiated by the farming community, in connection with the US Department of Agriculture, to reduce the impact of agricultural activities on water quality.

The main Beaver Creek channel and most of the tributaries were channelized in the early to mid-1900's to facilitate the drainage of land. Channelization has resulted in increased hydrologic responses such as rapidly rising water levels and increased stream velocity during rain events. In 1990, the U.S. Geological Survey initiated a water-quality monitoring research program. The program included collection and evaluation of water chemistry, stream hydrology and geomorphology data. Biomonitoring was added as a fourth parameter in early 1993 as part of this comprehensive project.

The modified hydrology and geomorphology of the BCW brought the applicability of established toxicity bioassessment protocols under question. It was unclear how the modified geomorphology and hydrology would affect the collection and interpretation of biomonitoring data with regard to chemical disturbance. Other ecological factors had to be considered as well, such as species distribution patterns in the different stream orders, geographic location of the watershed, seasonality and natural habitat. Thus, study were: (1) to evaluate and develop biomonitoring protocols appropriate for modified streams such as those in the BCW, (2) determine if the effects of chemical perturbation could be distinguished from hydrologic and geomorphic disturbances, and (3) characterize the general structure and function of a first-second order channelized stream in the BCW. This paper presents the results to the first objective and preliminary results of objective number 3.

METHODS AND MATERIALS

Macroinvertebrates ("organisms that are retained by a Standard #30 sieve [600 um mesh]") were selected as the indicator organisms because considerable research has demonstrated that they are excellent biological indicators of water quality (Klemm, et. al., 1990; Plafkin, et. al., 1989; Rosenberg and Resh, 1993). Benthic macroinvertebrates were collected with scoops and sieves during two reconnaissance trips through the basin. This was done to obtain a general census of the species distribution pattern in the basin. The difference in biological community structure between upstream and downstream sites indicated that if any correlation was to be made between macroinvertebrate response and chemical exposure, initial efforts should focus on a first and second order tributary. Therefore, the main monitoring sites were selected in a well characterized, channelized, first-second order tributary, draining a 300 hectare cotton and soybean field. The stream had intermittent riffles with perennial pools.

Sampling sites were chosen in this tributary to reflect unique riparian conditions. The riparian features represented were herbaceous bank vegetation and woody bank vegetation. The herbaceous and woody reaches of stream were each approximately 1 kilometer long and straight due to channelization. All the data reported herein were collected in

the pools of the tributary because flow in the riffle sections reduced to a trickle during the dry season. General site characteristics of the herbaceous and woody riparian stream segments are shown in Table 1.

TABLE 1. Site characteristics observed during June through September. [HRV, herbaceous riparian vegetation; WRV, woody riparian vegetation; FFB, forest flood basin; WR, wetland reference site.

		Sit	е	
Characteristics	HRV	WRV	FFB	WR
Lentic (ponded) water	perennial	perennial	perennial	perennial
Lotic (flowing) water	seasonal	seasonal	no	perennial
Temperature (°C) range	27-34	24-30	24-32	25-30
pH range	7.5-9.2	7.0-8.1	7.0-9.0	7.5-8.2
Width (meters) range	0.8-2.0	1.2-2.0	0.5-4.0	1.3-2.5
Depth (meters) range	0.1-0.5	0.1-0.7	.05-0.3	0.2-0.7
Periphyton (% coverage)	95	35	45	70
Woody debris in water	no	intermittent	yes	yes
Emergent aquatic plants	yes	no	no	yes

We were unsuscessful at locating a nonchannelized reference site with a similar hydrologic regime (flowing water during the wet season, and ponded water during the dry season) in the same watershed, so two reference sites were selected to represent these hydrologic conditions. The first reference site was in a flooded forest basin adjacent to the cotton field. This non-channelized site was annually inundated with 5 to 30 cm of ponded (lentic) water. The second reference site was located in another branch of Beaver Creek just downstream from a natural, hardwood, bottomland wetland and series of beaver ponds. This site was in a small third order stream with perennial flowing (lotic) water. The characteristics of the two non-channelized reference sites also are listed in Table 1.

Standard U.S. Environmental Protection Agency (EPA) biomonitoring protocols were reviewed and procedures were selected based on adaptability to the BCW system. Two conventional biomonitoring techniques, colonization of multiplate samplers and composition of the benthic macroinvertebrate community, were chosen from the EPA standard protocols (Klemm, et. al., 1990; Plafkin, et. al., 1989). The first procedure surveys the macroinvertebrates that colonize a Masonite, multiplate sampler placed in the water column and harvested after a set incubation time. This method has been standardized and comparisons are often made between studies in the literature (Rosenberg and Resh, 1993). The second method quantifies the macroinvertebrates that inhabit the top few inches of the stream bottom (LaPoint and Fairchild, Both methods are considered semi-quantitative because the number of organisms within a sampling area are counted. replicate samples per location (e.g., three multiplate samplers) were collected on each sampling date.

A less conventional biomonitoring method was also examined. The technique used natural material, such as leaf and cotton plant debris, as a substrate for colonization. The application of leaf packs as an indicator of biological integrity has been explored by other investigators (Elder and Cairns, 1982; Paul et al., 1978). The data gathered in those studies provided information on the function (i.e., biogeochemical processes) of the waterway. The present study modified and adapted the leaf-pack methods to include structure (e.g., quantify macroinvertebrate colonization) and function (e.g., rate of plant

debris decomposition). It was assumed leaf or cotton packs provided a better replication of substrates occurring in these streams for macroinvertebrate colonization than the artificial substrate-multiplate samplers. The leaf materials were collected in early winter from an adjacent bottomland forest, dried and sorted by genus. The composition of the leaves, dry weight (in grams), and percent of sample per bag were: oak 7.5 (50%), willow 2 (13%), hickory 1.5 (10%), sweet gum 1 (6.6%), silver maple 1 (6.6%), cottonwood 1 (6.6%) and dogwood 1 (6.6%). Cotton stems and bolls ("cotton debris") were also collected, dried, weighed, and placed in similar mesh bags. Cotton was selected because it is a primary crop in the Beaver Creek basin and large quantities of debris have been reported to wash into receiving streams after harvest during intense rain events.

The plant material was dried in an oven at 70°C until the decrease in dry weight stopped (about 1 week). Predetermined dry weights of plant material (15.0 g leaves, 35 g cotton) were placed in labelled 1.6 mm mesh bags of 10 by 10 cm size. The bags were closed and tied off to 2-ft stakes with nylon string. They were installed in late April 1993 at the same time as the multiplate samplers. Replicate samples were retrieved in early June, early July and early September. The contents of the bags were emptied into plastic tubs and the macroinvertebrates were sorted out, identified, quantified and preserved for identity confirmation. The plant material was gently cleaned with deionized water and dried in an oven at 70°C until dry. The resulting weight was recorded and the change in weight was calculated for each bag of plant material.

Biological samples were collected 6, 12 and 18 weeks after the substrates were placed in the field. The substrate samplers were enclosed with zip-lock plastic bags and placed on ice until the organisms could be harvested, taken to the laboratory, and preserved in 70% alcohol. The benthic sediment samples were collected using a long handled shovel with a special steel scoop, 8 inches across, 10 inches long and having 2 inch sides. The collected sediment was sifted in the field and the macroinvertebrates were preserved in 70% alcohol, brought to the laboratory and identified to Family. Surface-water samples were collected every 5 minutes during storm flow at the water-chemistry sampling station separating the herbaceous and woody riparian zones. These samples were analyzed for pesticides, nitrogen and sediment load. Preliminary results of the chemical analyses indicate that pesticides were not a problem during this study period. Additional field measurements were made every 2 to 3 weeks during the study period to monitor low-flow conditions of water depth, pH, temperature, and dissolved oxygen. Aquatic plant and animal species observations were also recorded at this time. Many of these organisms represent sentinel indicator species analogous to macroinvertebrates (Table 1).

Data observed or calculated for this study included abundance (total number of organisms), taxonomic richness (number of Families), mean diversity of taxon (Shannon-Weaver mean diversity, Klemm, et al., 1990) and Beck's biotic index (Beck, 1955; Terrell and Perfetti, 1989; Resh and Jackson, 1993). The data are reported on a temporal and spatial comparison, and the average change in dry weight of replicate leaf and cotton debris packs are reported as a function of time. Standard deviations for the Beck biotic index are reported.

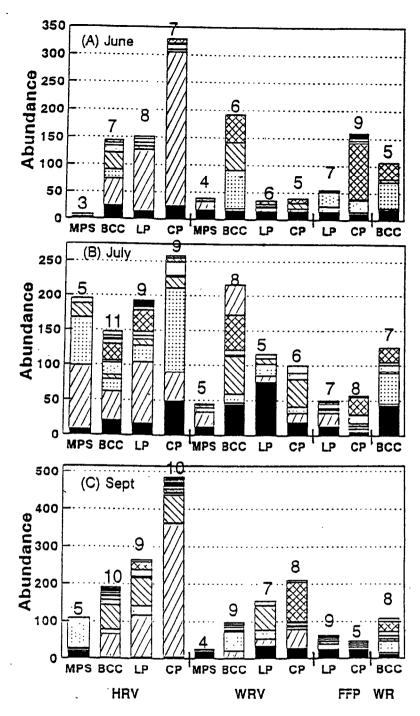
RESULTS

Community structure estimated by the abundance of macroinverte-brates at each location and sampling date is depicted by the height of the bars in Figure 1a-1c. Abundance is the total number of organisms counted per 3 replicate samples at each site. The overall abundance of organisms collected increased over the duration of the study (from 1259 in June, to 1498 in July and 1651 in September). The multiplate samplers generally had the lowest abundance of the four sampling methods tested. The cotton packs located in the herbaceous riparian vegetation streams always provided the greatest abundance on each sampling date. However, cotton packs located at the other sampling locations did not have the same trend. Benthic community sampling appeared to be the most consistent sampling technique with regard to abundance at each location or time.

TABLE 2. Richness, abundance, and mean diversity of sampling method. [HRV, herbaceous riparian vegetation; WRV, woody riparian vegetation; FFB, forest flood basin; WR, wetland reference site; MPS, multiplate sampler; BCC, benthic community composition; LP, leaf pack; CP, cotton pack; R, richness, number of Families; Ab, abundance, sum total; MD, mean diversity; ---, no data collected]

			MPS		BC	C		LP			CP	
Site	R		MD	R	Ab	MD	R	Ab	MD	R	Ab	MD
June												
HRV	3	8	1.36	7	144	2.49	8	151	1.36	7	329	0.87
WRV	4	39	1.52	6	192	1.95	6	35	2.10	5	39	2.55
FFB							7	56	2.09	9	161	1.77
WR				5	105	1.73						
July												
HRV	5	195	1.75	10	145	3.73	9	194	2.50	9	258	2.57
WRV	5	45	1.92	8	216	2.46	5	116	1.58	6	100	2.34
FFB							8	50	2.35	8	57	2.41
WR				7	126	2.18						
Sept												
HRV	5	108	1.85	10	144	2.11	9	265	2.20	9	475	1.26
WRV	4	25	1.38	9	98	2.09	7	132	1.93	8	209	2.01
FFB							9	77	2.73	5	46	1.99
WR				8	108	2.69						
Avg*	4.	5 70	1.63	8.3	3 151	2.47	7.3	3 149	1.95	7.3	3 235	1.93
*Avera	ages	calcu	ılated us	ing H	RV and	WRV val	ues o	nly fi	com all t	hree	sampl	ing dates

Richness is a measure of the number of different taxa (Families) found at each location and sampling date. The richness of each sampling technique during each sampling date is depicted in Figure 1 and tabulated in Table 2. The patterns shown in the bars of Figure 1 represent different taxa (Families) collected from each site. The height of each pattern represents the number of organisms within that taxon. In general, the benthic community sampling provided the greatest measure of richness compared to the three substrate-sampling techniques (multiplate samplers, leaf packs and cotton packs). Multiplate samplers consistently gathered the poorest taxon-richness of the 4 techniques at any particular site.



(# on top of bar = # of Families)

Figure 1. Richness and diversity as measured by sampling technique collected in (a) June (b) July and (c) September 1993. Patterns in the bars indicate abundance of different taxa. [HRV, herbaceous riparian vegetation; WRV, woody riparian vegetation; FF3, forest flood basin; WR, wetland reference site; MPS, multiplate sampler; BCC, benthic community composition; LP, leaf pack; CP, cotton pack]

Mean diversity is a measure of taxonomic diversity and distribution. The scale increases with increasing diversity and distribution of organisms between the taxa. The rationale behind this measure is that species diversity decreases with decreasing water quality. Quite frequently a particular Family dominated the percentage of organisms collected using a particular sampling technique or date.

For example, a high density of organisms was observed in the cotton packs collected from the herbaceous riparian stream in June (Fig. 1). Yet, only one species of snail represented almost 90% of the total number of organisms. This resulted in a low mean diversity (Table 2). Of the four sampling techniques tested, the benthic community grab samples provided the highest value for average mean diversity. Smaller values were associated with leaf packs and cotton packs, and the smallest values were provided by multiplate samplers (Table 2).

Beck's biotic index is a score that weighs the taxonomic richness of the sample and the tolerance of the taxa to pollution. Organisms are classified by three categories: sensitive, moderately sensitive and tolerant of pollution. The categories for Beck's biotic index were established from ecological surveys in Florida (Resh and Jackson, 1993). Points are given for sensitive and moderately sensitive taxa. A score of 10 or less is considered polluted water and warrants further attention to determine and remediate the problem (Terrell and Perfetti, 1989). With one exception, the multiplate samplers always had a very low biotic index score that indicated a polluted stream (Table 3). The benthic community grab samples consistently resulted in healthy scores. Thus, one could reach completely opposite conclusions from the same location in the BCW depending on the sampling technique used. The leaf pack and cotton packs each averaged moderately healthy biotic index scores over time and location. These data suggest that streams where physical habitat is lacking or constantly changing, or where predation is highly selective, biotic indices scores are tremendously influenced by the sampling technique used.

Table 3. Beck's biotic index for the four sampling techniques. [HRV, herbaceous riparian vegetation; WRV, woody riparian vegetation; FFB, forest flood basin; WR, wetland reference site; MPS, multiplate sampler; BCC, benthic community composition; LP, leaf pack; CP, cotton pack; ---, no data;] The bottom line [CV] is the coefficient of variation of Beck's biotic index for the four sampling techniques at the HRV and WRV sites.

the nkv	and wav sice			
Site	MPS	BCC	LP	CP
June				
HRV	0	71	25	18
WRV	2	122	14	11
FFB			40	37
WR		54		
July				
HRV	89	59	78	44
WRV	10	75	35	57
FFB			32	24
WR		59		
Sept				
HRV	10	45	104	94
WRV	8	64	67	123
FFB			47	18
WR		50		
CV	1.6	0.31	0.54	0.76

A basic stream function was evaltuated by measuring the change in leaf weight over time at three of the sites. This was done to determine how rapidly plant material would decompose under stream

conditions in the BCW. The results are shown in Figure 2. leaf material and the cotton debris lost approximately 35% of their initial dry weight over the course of the study. The material in the forest flood basin location was slowest to lose weight. In fact, the leaves gained weight due to sediment embedding in the material. Sediment embedded in leaf material at the other sites as well, but the amount on the leaves in the forest flood basin was greatest. Forest flood plains have been noted for their ability to retain sediment (Mattraw and Elder, 1984). It has also been demonstrated that plant material will usually decompose faster under permanently inundated conditions (Elder and Cairns, 1982). The cotton debris was less vulnerable to embedding sediment material and lost much of its weight in the first 6 weeks. This quick weight loss could be explained by a quick loss in easily consumable food, followed by a slow decay in the less energy efficient fibers and waxes. Microscopic examination of the plant material revealed that fungal hyphae were more prominent on the cotton stalks than leaf material. The leaf material sustained macroinvertebrates classified as shredders (Cummins, 1973) frequently than the cotton material. However, leaf material that was heavily embedded with sediment particles had noticeably fewer shredders than particle-free material.

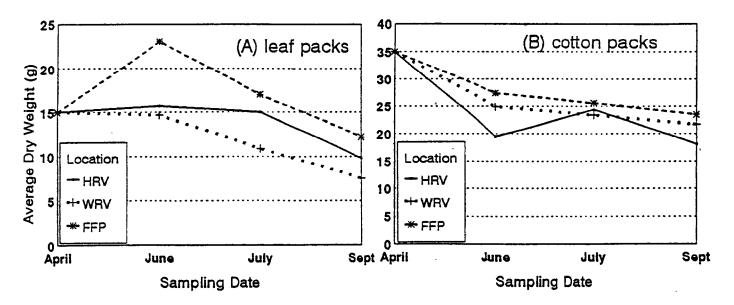


Figure 2. Average weight of the (A) forest leaves and (B) cotton debris as a function of time at three sites.

DISCUSSION AND CONCLUSION

There were distinct differences between the sampling methods used and the results of the biological assesment. The benthic community grab sampling technique provided the best scores for mean diversity and Beck's biotic index compared to the use of plant-material packs or artificial substrates. In fact, the results for multiplate samplers and benthic community grab samples were completely opposite. The multiplate samplers failed to attract many species even after an 18 weeks incubation period. Multiplate samplers did not provide a true picture of the taxonomic richness or abundance in this particular

tributary.

The results support the assertion by LaPoint and Fairchild (1992) that benthic grab sampling provides the best representation of the benthic community composition and ecolological health of the system. The consistent scores and low coefficient of variation (Table 3) resulting from the use of this method supports this hypothesis. Proper sampling of the benthos precludes many of the sampling biases associated with substrate colonization studies. Benthic grab sampling requires more time than artificial substrates, but the data appear to be more accurate.

The assumption behind substrate sampling is that it removes habitat as a variable and provides standardized surface between sites. The plant material packs, which served as more natural substrates, had greater macroinvertebrate taxonomic richness and biotic index scores than the multiplate substrates. Some of this could be explained by the fact that the leaf and cotton material served as a primary or secondary food source as well as a shelter for the organisms. The low mean diversity in several of the cotton- and leaf-pack samplers could be explained by the removal of predation pressure. For example, 329 macroinvertebrates were counted in the cotton packs collected in June from the herbaceous riparian location. Eighty-eight percent of these organisms were snails that could move about freely without fear of predation. Snail-eating beetles, common in that reach of the tributary, were excluded from the sample because they were unable to get through the mesh. Therefore, the leaf and cotton debris packs biased the outcome of the stream survey.

Channelizing a stream results in increased water velocity and a reduction in the debris retention-time, producing a flushing effect (Petersen and Petersen, 1991). Inspection of the tributary revealed that most of the plant debris moved a significant distance with each rain event. Thus, very little information about the rate of decomposition or the ability of the system to recycle this material has been determined. Retaining plant debris in packs provided some information on the function of the stream. Results confirm that many of the leaves in the forest leaf packs would decompose at a steady rate after an initial weight gain (Figure 2). The cotton debris however, appeared to be resistent to decomposition after the initial weight loss. This indicates that the stream could process and transform forest leaf-material faster than cotton crop debris.

In summary, based on the results of the different sampling strategies tested, multiplate samplers were found inadequate for biomonitoring a channelized stream with conditions similar to the BCW. The data derived from multiplate samplers were misleading. The plant material packs were valuable in determining the functions of the stream. However, they had some drawbacks when used to characterize stream community structure. Benthic grab sampling was the most effective sampling method tested in this study. Plant material packs and benthic grab sampling each have their advantage. They should be considered jointly when designing a biomonitoring project for channelized streams such as in the Beaver Creek Watershed.

REFERENCES

Beck, W.M., Jr. 1955. Suggested Method for Reporting Biotic Data. Sewage and Industrial Wastes 27:1193-1197.

- Cooper, C.M., 1993. Biological Effects of Agriculturally Derived Surface Water Pollutants on Aquatic Systems a review. *J. Environ. Qual.* 22:402-408.
- Cummin, K.W., 1973. Trophic Relations of Aquatic Insects. Ann. Review of Entomology 18:183-206.
- Elder, J.F. and D.J. Cairns, 1982. Production and Decomposition of Forest Litter Fall on the Apalachicola River Flood Plain, Florida. U.S. Geological Survey Water-Supply Paper 2196-B, pp B12 B32.
- Klemm, D.J., P.A. Lewis, F. Fulk and J.M. Lazorchak, 1990.

 Macroinvertebrate Field and Laboratory Methods for Evaluating the
 Biological Integrity of Surface Waters. EPA/600/4-90/030, pp 109-158.
- LaPoint, T.W. and J.F. Fairchild, 1992. Evaluation of Sediment Contaminant Toxicity: The Use of Freshwater Community Structure. in Sediment Toxicity Assessment, ed. G.A.Burton. Lewis Publishing, Ann Arbor, MI, pp. 87-110.
- Mattraw, H.C., Jr. and J.F. Elder, 1984. Nutrient and Detritus Transport in the Apalachicola River, Florida. U.S. Geological Survey Water-Supply Paper 2196-C, pp. C31-C55.
- Paul, R.K., Jr., E.F. Benfield and J.D. Cairns, Jr., 1978. Effects of Thermal Discharge on Leaf Decomposition in a River Ecosystem. Verh. Internat. Verein. Limnol. 20:1759-1766.
- Petersen, L.B.M. and R.C. Petersen, Jr., 1991. Short term retention properties of channelized and natural streams. Verh. Internat. Verein. Limnol. 24:1756-1759.
- Plafkin, J.L., M.T. Barbour, K.D. Porter, S.K. Gross and R.M. Hughes, 1989. Rapid Bioassessment Protocols for use in Streams and Rivers: Benthic Macroinvertebrates and Fish. EPA/440/4-89/001.
- Resh, V.H. and J.K. Jackson, 1993. Rapid Assessment Approaches to Biomonitoring Using Benthic Macroinvertebrates, in Freshwater Biomonitoring and Benthic Macroinvertebrates, eds. D.M. Rosenberg & V.H. Resh. Chapman and Hall, New York, N.Y., pp. 195-233.
- Rosenberg, D.M. and V.H. Resh, 1993. Introduction to Freshwater Biomonitoring and Benthic Macroinvertebrates. *in* Freshwater Biomonitoring and Benthic Macroinvertebrates, eds. D.M. Rosenberg & V.H. Resh, Chapman and Hall, New York, N.Y., pp 1-9.
- Terrell, C.R. and P.B. Perfetti, 1989. Water Quality Indicators Guide: Surface Waters. U.S. Department of Agriculture, Soil Conservation Service, Washington, D.C., pp. 43-68.
- U.S. Environmental Protection Agency, 1989. Report to Congress; Water Quality of the Nation's Lakes. EPA Rep. 440/5-89-003. Office of Non-point Source Control Branch, Washington, DC.
- U.S. Environmental Protection Agency, 1992. Framework for Ecological Risk Assessment. EPA630/R-92/001. Risk Assess. Forum, Washington, DC.

TRANSPORT AND DEGRADATION OF ALDICARB IN THE SOIL PROFILE: A COMPARISON OF CONVENTIONAL TILLAGE AND NON-TILLAGE

Lisa D. Olsen¹, Angel Roman-Mas², Carol P. Weisskopf³, and Stephen J. Klaine³

ABSTRACT: Paired cotton fields (conventionally tilled and non-tilled) in the Beaver Creek watershed, West Tennessee, were selected to (1) develop a sampling strategy that accurately characterizes the spatial and temporal distribution of aldicarb and its metabolites in the soil profile, (2) evaluate the transport and degradation of aldicarb and its metabolites in the soil profile, and (3) compare the behavior of aldicarb and its metabolites in conventionally tilled and non-tilled cotton fields. The observed concentration distributions support sampling in multiple positions within transects perpendicular to the orientation of the rows. Multiple transects were proven necessary to achieve field representation for a given transect position. This sampling strategy provided suitable data sets for calculating representative and statistically defensible field values. Base-catalyzed and surface-catalyzed hydrolysis appeared to be the most important mechanisms for the disappearance of aldicarb sulfoxide in the soil profile in both tillage systems. Degradation rate constants were 0.046 day⁻¹ for the conventionally tilled field. The half-lives of aldicarb sulfoxide were 15 days and 16 days for the conventionally tilled and the non-tilled fields, respectively. Horizontal transport was negligible, and vertical transport was minimal for both tillage systems.

KEY TERMS: transport; degradation; soils; aldicarb; aldicarb sulfoxide; aldicarb sulfone.

INTRODUCTION

Non-tillage has been effectively used to reduce soil erosion. However, researchers have suggested that non-tillage can enhance chemical transport and increase the potential for ground-water contamination (Dick and others, 1989; Hall and others, 1989; Isensee and others, 1990). Resource-management agencies in the Mississippi embayment, where the soils are highly susceptible to erosion, have recommended non-tillage as a best management practice (BMP). The risk of ground-water contamination associated with the implementation of non-tillage in this region needs to be addressed before this BMP is widely adopted.

Field studies comparing pesticide transport and degradation in conventionally tilled and non-tilled soils in this region are needed. An important precursor for such studies is the development of sampling strategies that systematically address the spatial and temporal distribution of pesticides within the soil profile. An insufficient characterization of the pesticide distribution can bias the evaluation of pesticide transport and degradation, and subsequently can bias the comparison of conventional tillage and non-tillage results.

^{&#}x27;Graduate Student, Department of Geography, University of Memphis, Memphis, TN 38152.

²Hydrologist, U.S. Geological Survey, 7777 Walnut Grove Rd., Suite LLB-2, Memphis, TN 38120.

^{&#}x27;Assistant Professor and Associate Professor, respectively, The Institute of Wildlife and Environmental Toxicology, Clemson University, Pendleton, SC 29670.

In 1993 the U.S. Geological Survey, in cooperation with the Tennessee Department of Agriculture, initiated a study was initiated to (1) develop a sampling strategy that accurately characterizes the spatial and temporal distribution of aldicarb and its metabolites in the soil profile, (2) evaluate the transport and degradation of aldicarb and its metabolites in the soil profile, and (3) compare the behavior of aldicarb and its metabolites in conventionally tilled and non-tilled cotton fields. This study is part of a comprehensive research and monitoring program to evaluate agricultural non-point source pollution and BMP's in the Beaver Creek watershed, West Tennessee. This report discusses results for the 1993 growing season.

ALDICARB DEGRADATION

Aldicarb is a widely used insecticide and nematicide for cotton crops in the Beaver Creek watershed. Granular aldicarb is typically integrated into the soil during planting. The 0.3 meter (m) wide application band is centered along the row tops. Application rates range from 0.6 to 0.9 kilogram per hectare (kg/ha) of the active ingredient. Upon contact with soil moisture, aldicarb is released. Plant uptake is generally less than 5 percent of that applied (Smith and Parrish, 1993). Transport in surface runoff is less than 10 percent of that applied.

Because of its high water solubility (5,730 milligrams per liter), aldicarb is considered highly mobile (Seiber, 1987). However, aldicarb mobilization can be retarded by adsorption into organic phases in the soil profile. Rao and others (1986) reported that in sandy soils, aldicarb sorption is controlled by the organic carbon content. The molecular structure of aldicarb implies that the pesticide has potential for binding ionically to clays. However, the sorption mechanism and sorption coefficient are unknown.

Laboratory experiments conducted by Lightfoot and others (1987) show that microbial oxidation is the most important degradation mechanism for aldicarb in soils. Half-lives of 1 to 2 days were reported in these experiments. The first product of aldicarb oxidation is aldicarb sulfoxide, which can further oxidize to form aldicarb sulfone. Aldicarb, aldicarb sulfoxide, and aldicarb sulfone concurrently hydrolyze to form non-carbamate oximes and nitriles.

Hydrolysis is the most important degradation mechanism for aldicarb sulfoxide and aldicarb sulfone in soils (Lightfoot and others, 1987; Hansen and Speigel, 1983; Lemeley and Zhong, 1984). Data presented by Lightfoot and others (1987) show that the hydrolysis rates for aldicarb sulfoxide and aldicarb sulfone follow first-order kinetics and are pH and temperature dependent. Lightfoot and others (1987) also reported half-lives for these metabolites in experimental soil microcosms ranging from 10 days to greater than 2,000 days, depending upon the pH and temperature.

Aldicarb and its metabolites are relatively toxic (Fukuto, 1987). Their primary mechanism of action is to inactivate the acetylcholinesterase enzyme (Fukuto, 1987; Metcalf, 1971). Rat oral LD₅₀ values are 1 milligram per kilogram (mg/kg) for aldicarb and aldicarb sulfoxide, and 24 mg/kg for aldicarb sulfone (Lightfoot and others, 1987). Non-carbamate metabolites of aldicarb are of little environmental concern because their toxicities are relatively low and they continue to degrade to form aldehydes, acids, and alcohols.

PROTOCOLS AND METHODS

Two adjacent cotton fields in the Beaver Creek watershed were selected for this study (See Figure 1). The fields are located in Shelby County, near the Collierville-Arlington Road. These fields are of similar size, slope, and soil type. The south field was cultivated by conventional tillage (the top 0.3 meter was disked before planting). The north field was cultivated by non-tillage (crop residues from the previous growing season were not disturbed during planting). The nominal rate of application was 0.84 kg of aldicarb (active ingredient) per hectare. Agricultural limestone was added to these fields as recommended by the University of Tennessee, Agricultural Extension Service (John C. Wilson, farmer, oral commun., 1993).

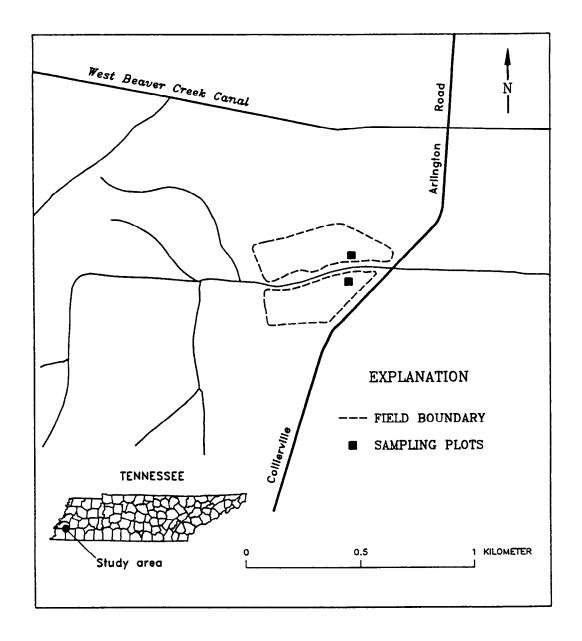


Figure 1. Location of study area and sampling plots.

Soils in these fields were classified in 1966 by the U.S. Department of Agriculture, Soil Conservation Service, as Falaya silt loam. Soil pH ranges from 5.1 to 5.5. However, these soils are usually limed to bring the pH to about 6.5. The water table is often within a foot of the land surface in winter and spring, and declines several feet below the surface in the summer and fall (U.S. Department of Agriculture, Soil Conservation Service, 1989).

Sampling Protocols

A 30- by 30-m sampling plot was delineated in each field. All sampling in this study was conducted within the plot boundaries. Prior to pesticide application, soil samples were taken from these plots to verify the absence of aldicarb and its metabolites. Aldicarb was applied to both fields on May 18, 1993. Soil samples were collected from the conventionally tilled field at 9, 22, 52, 84, 115, and 149 days after the application of aldicarb and from the non-tilled field at 3, 21, 43, 70, 113, and 148 days after the application date. For each sampling

date, three transects were randomly established from row top to adjacent row top (see Figure 2). Five equidistant coring positions were sampled in each transect. Soil cores were taken at each position in 0.15-m intervals from the land surface to a maximum depth of 0.90 m. Soil cores were collected using a 0.054-m diameter stainless-steel auger. The auger was cleaned with deionized water between sampling depths to avoid contamination from one depth interval to the next. For the same reason, the top 3 centimeters of each soil core (except for the uppermost interval) were discarded.

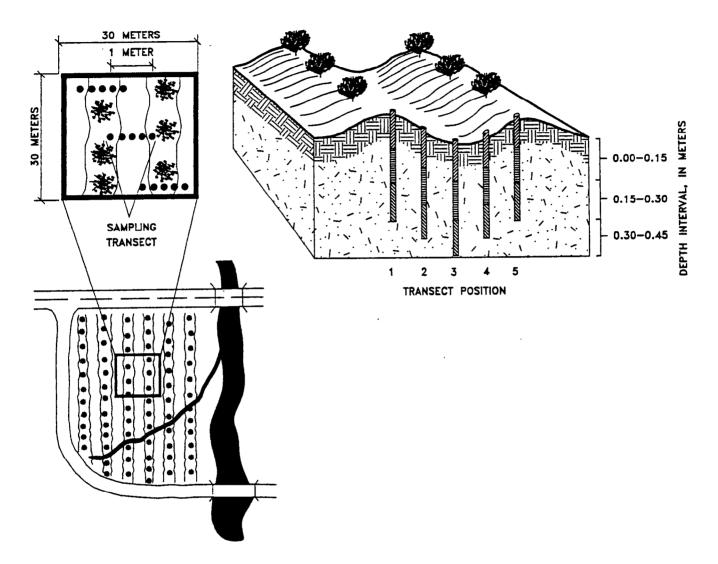


Figure 2. Schematic diagram of soil-sampling strategy.

Each soil core (0.15 m long) was transferred to a large sealable plastic bag and homogenized. Half of each soil sample was transferred to a second plastic bag, packed with ice, and sent by overnight carrier to The Institute of Wildlife and Environmental Toxicology (TIWET), Clemson University, for analysis of aldicarb, aldicarb sulfoxide, and aldicarb sulfone.

Analytical Methods

The samples were analyzed using TIWET standard operational procedure #401-53-01. To summarize the procedure, aldicarb residues for each sample were isolated by shaking together 0.050 kg of homogenized soil

and 0.100 kg deionized water in a sealed polyethylene bottle for 1 hour, centrifuging the slurry, and extracting the supernatant with methylene chloride. The methylene chloride fractions were then rotary evaporated to dryness. Aldicarb residues for each sample were resuspended in 2.0 mL of acetonitrile and analyzed with a Hewlett-Packard 1090 Liquid Chromatograph (any use of trade, product, or firm names in this publication is for descriptive purposes only and does not imply endorsement by the U.S. Government) with a fluorescence detector.

Ten percent of the samples were run in triplicate. Analyses were accepted only if the coefficients of variation were less than 5 percent. Laboratory blanks representing 5 percent of the total number of samples were also analyzed.

RESULTS AND DISCUSSION

Most of the results and discussion presented in this paper are centered on the transport and degradation of aldicarb sulfoxide. Aldicarb was not detected after the first sampling date in each field, and the occurrence and distribution of aldicarb sulfone was similar to that of aldicarb sulfoxide.

Evaluation of Sampling Protocols

In general, the occurrence of aldicarb sulfoxide was limited to the row tops (transect positions 1 and 5) where the pesticide was applied (see Figure 2). Aldicarb sulfoxide concentrations in the slopes (positions 2 and 4) and furrows (position 3) were low or below the detection limit of 1.5 micrograms per kilogram. This distribution was observed in all transects, at all depth intervals, for all sampling dates, and in both fields (see Figure 3).

For a given sampling date, the variances in the aldicarb sulfoxide concentrations at the row-top position were relatively high among and within transects (see Table 1). Therefore, a larger number of samples are needed to obtain representative concentrations at this position than at the slope and furrow positions where concentration variances were smaller.

The aldicarb sulfoxide concentration distribution supports sampling in multiple positions within transects perpendicular to the orientation of the rows. Because the concentrations for a given transect position varied widely among transects, multiple transects proved necessary to achieve field representation for that particular position. Considerable spatial variability in the pesticide residue concentrations in soil samples has been reported in many field studies (Smith and others, 1987; Jones and others, 1988; Hornsby and others, 1990). Concentration medians were assumed to provide representative and statistically defensible field values for a given transect position. Medians were selected over other measurements of central tendency because they are less biased by extreme values and, therefore, better suited to describe data sets with large variances. Concentration medians were used to characterize the spatial and temporal distribution of aldicarb sulfoxide in the two fields.

Transport

Aldicarb sulfoxide concentration distributions were used to determine the vertical and horizontal transport in the soil profile. Because aldicarb was applied at the row top position at about 0.07 m below land surface, occurrence of aldicarb or its metabolites in other transect positions or depths would constitute horizontal or vertical transport. The non-occurrence of aldicarb sulfoxide in the slope and furrow positions of both fields indicates that horizontal transport is negligible. The absence of horizontal mobilization for aldicarb and its metabolites was also observed by Smith and Parrish (1993) in peanut fields in sandy loam soils.

Aldicarb sulfoxide concentrations peaked in the uppermost interval, 0 to 0.15 m, throughout the sampling period (See Figure 4 and Table 1). Concentrations in the uppermost interval of the row-top position accounted for about 85 percent of the total aldicarb sulfoxide in the soil profile. No occurrence of aldicarb sulfoxide was

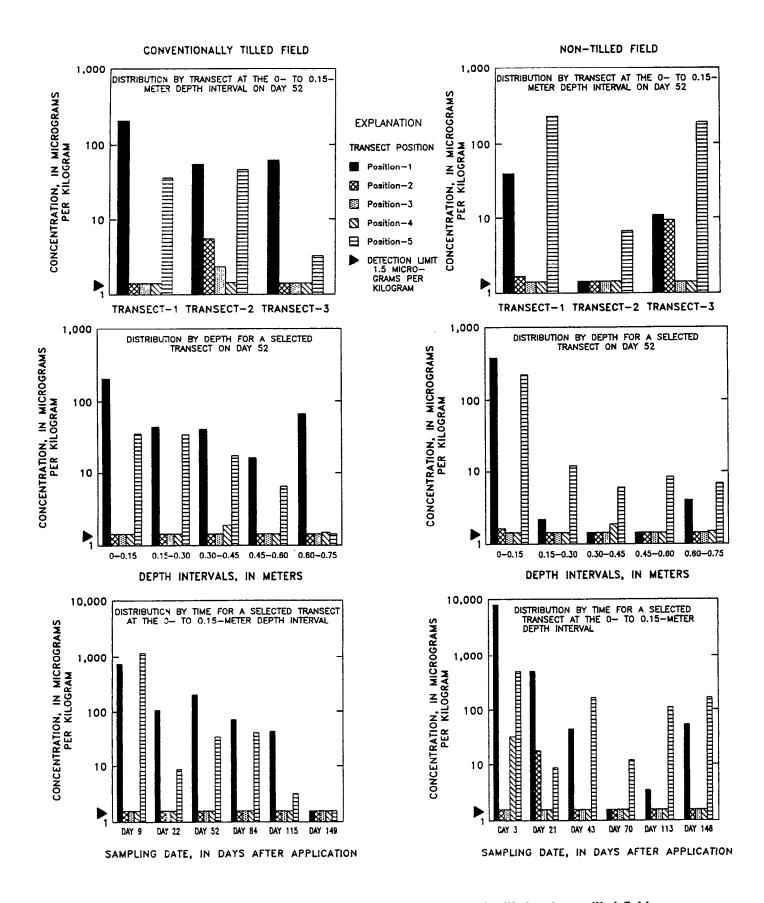


Figure 3. Typical aldicarb sulfoxide distribution for the conventionally tilled and non-tilled fields.

Table 1.-Descriptive statistics for aldicarb sulfoxide in the conventionally tilled and non-tilled fields

[m, meters; n, number of observations; µg/kg, micrograms per kilogram; (µg/kg)², micrograms per kilogram squared; <, less than; -, no data are available]

Conventionally silled	Ω	Day 9	Day	Day 22	Day	Day 52	Day	Day 84	Day	Day 115	Da	Day 149
Conventionally filled	Median (µg/kg)	Median Variance (µg/kg) (µg/kg)²	Median (µg/kg)	Median Variance (μg/kg) (μg/kg)²	Median (µg/kg)	Median Variance (μg/kg) (μg/kg) ³	Median (μg/kg)	Median Variance (μg/kg) (μg/kg) ³	Median (µg/kg)	Median Variance (µg/kg) (µg/kg)²	Median (ug/kg)	Median Variance (µg/kg) (µg/kg) ²
0 to 0.15 m												
Row top, n=6	1,005	87,616		1,901	50.5	5,213	16.5	759	3.9	320	<1.5	0.0
Slope, n=6	<1.5	5.4	<1.5	0.0	<1.5	2.8	<1.5	0.0	<1.5	0.0	<1.5	0.0
Furrow, n=3	9.9	23.1		1,030	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
0.15 to 0.30 m												
Row top, n=6	25.5	3,003	2.2	610	2.8	1,089	2.4	2.3	<1.5	0.5	<1.5	0.0
Slope, n=6	<1.5	7.5	<1.5	9.0	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
Furrow, n=3	3.4	2.2	<1.5	0.0	<1.5	0.3	<1.5	0.0	<1.5	0.0	<1.5	0.0
0.30 to 0.45 m												
Row top, n=6	79.0	5,432		1,544	30.0	1,183	2.0	0.7	<1.5	0.1	<1.5	0.0
Slope, n=6	<1.5	64.3	<1.5	0.0	<1.5	0.5	<1.5	0.0	<1.5	0.0	<1.5	0.0
Furrow, n=3	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
0.45 to 0.60 m												
Row top, n=6	1	t	ı	ı	<1.5	396	<1.5	0.0	<1.5	0.0	<1.5	0.0
Slope, n=6	ı	ł	i	ŀ	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
Furrow, n=3	:	ł	ı	1	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
0.60 to 0.75 m												
Row top, $n=6$	ı	!	ı	ı	7.9	681	<1.5	0.0	<1.5	0.0	<1.5	0.0
Slope, n=6	ı	ŀ	i	:	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
Furrow, n=3	ŀ	ŀ	ı	ı	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
0.75 to 0.90 m												
Row top, n=6	1	1	ı	ı	:	1	1	ı	<1.5	0.0	<1.5	0.0
Slope, n=6	ı	ı	ŧ	ı	ı	:	ı	ı	<1.5	7.0	<1.5	0.0
Furrow, n=3	ı	1	1	1	1	t	ı	i	<1.5	0.0	<1.5	0.0

Table 1.-Descriptive statistics for aldicarb sulfoxide in the conventionally tilled and non-tilled fields-Continued

Modian Variance Modian Modian Variance Modian Variance Modian Modia	Mon Filled	Ď	Day 3	Da	Day 21	Da	Day 43	Da	Day 70	Day	Day 113	Day	Day 148
n=6 280 67,600 90.5 4816 9.7 2,272 24.5 10,201 2.0 180 30.0 61.5 n=6 4.7 22.6 <1.5	170ii-tiiled	Median (µg/kg)		Median (μg/kg)	Variance (μg/kg)³	Median (μg/kg)	Variance (μg/kg)³	Median (μg/kg)	Variance (μg/kg)²	Median (μg/kg)	Variance (μg/kg)³	Median (μg/kg)	Variance (μg/kg)²
15 15 15 15 15 15 15 15	0 to 0.15 m												
15	Row top, n=6		57,600		4,816		2,272		10,201	2.0	180	30.0	615
150 449 1.6 2.2 2.5 166 1.8 18.1 1.15 9.1 3.6 1.8 1.	Slope, $n=6$	4.7	22.6	<1.5	19.5	<1.5	1.1	<1.5	9.5	<1.5	0.0	<1.5	0.0
n=6 15.0 449 1.6 2.2 2.5 166 1.8 18.1 <1.5	Furrow, n=3	<1.5	0.0	<1.5	3.2	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
n=6 150 449 1.6 2.2 2.5 166 1.8 18.1 <1.5 9.1 3.6 n=6 <1.5 0.1 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 </td <td>0.15 to 0.30 m</td> <td></td>	0.15 to 0.30 m												
1	Row top, n=6	15.0	449	1.6	2.2	2.5	166	1.8	18.1	<1.5	9.1	3.6	8.5
n=3 <1.5 6.8 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.1 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5	Slope, n=6	<1.5	0.1	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
n=6 5.1 32.9 <1.5	Furrow, n=3	<1.5	8.9	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
n=6 5.1 32.9 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <1.5 <	0.30 to 0.45 m												
n=6 <1.5	Row top, n=6	5.1	32.9	<1.5	172	<1.5	106	<1.5	5.3	<1.5	1.5	<1.5	1.3
n=3 <1.5 0.7 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5 0.0	Slope, n=6	<1.5	0.0	<1.5	15.4	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
n=6	Furrow, n=3	<1.5	0.7	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
n=6 <1.5	0.45 to 0.60 m												
n=6 -	Row top, n=6	;	1	i	!	<1.5	9.4	<1.5	237	<1.5	0.0	<1.5	0.1
n=3 <1.5 0.0 <1.5 0.0 <1.5 0.0 <1.5	Slope, n=6	i	1	1	1	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
n=6 <1.5	Furrow, n=3	I	;	1	:	<1.5	0.0	<1.5	0.0	<1.5	0.0	<1.5	0.0
n=6	0.60 to 0.75 m												
n=6	Row top, n=6	ï	i	i	:	<1.5	0.0	1.8	8.4	<1.5	0.0	<1.5	0.0
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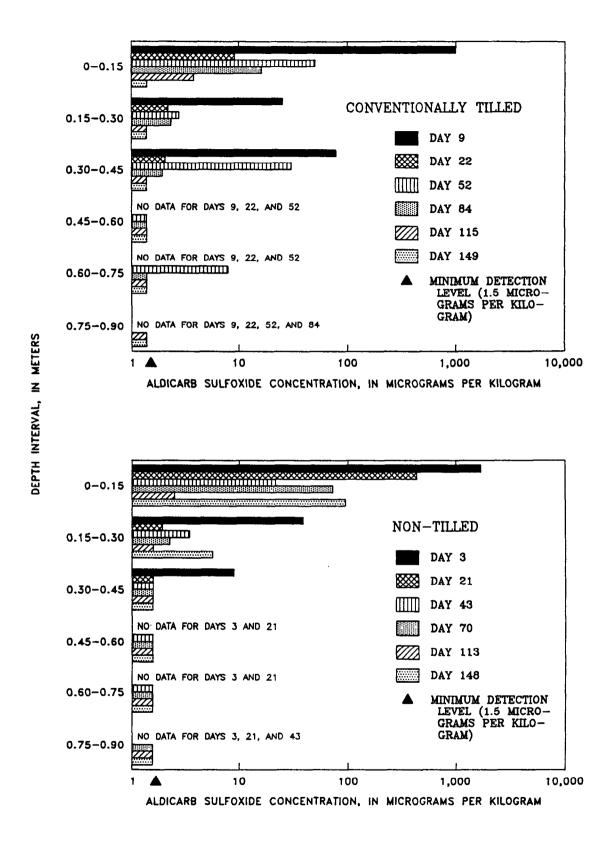


Figure 4. Median aldicarb sulfoxide for row-top position.

observed below the 0.60- to 0.75-m depth interval in either field. This indicates that the vertical transport in the row-top position was minimal. However, it was not determined if aldicarb sulfoxide had migrated below the 0.30- to 0.45-m depth interval early in the growing season. Soil samples below this depth interval were not collected until day 53 and day 43 in the conventionally tilled and the non-tilled fields, respectively.

Degradation

The median aldicarb sulfoxide concentrations for the 0 to 0.15-m depth interval were observed to decrease exponentially with time (see Figure 5). Correlation coefficients for the natural logs of the median aldicarb sulfoxide concentrations and time since application were -0.80 and -0.61 for the conventionally tilled and the non-tilled fields, respectively (See Figure 5). An analysis of the residuals showed that the data points for day 22 in the conventionally tilled field and for day 148 in the non-tilled field are outliers, therefore, respective adjusted correlation coefficients were -0.99 and -0.92. These correlation coefficients demonstrate that the disappearance of aldicarb sulfoxide at the 0 to 0.15-m depth interval follows a first-order kinetics reaction.

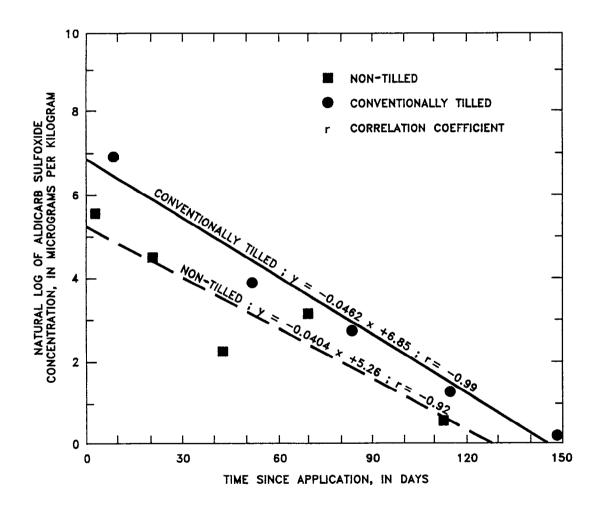


Figure 5. Natural log of the median aldicarb sulfoxide concentration for soil samples at the 0- to 0.15-meter depth interval within the band of application as a function of time since application.

The degradation rate constant, k, is the negative of the slope of the line generated by the correlation of the natural log of aldicarb sulfoxide concentration and the time since application (see Figure 5) (Castellan, 1983). Degradation rate constants were 0.046 day⁻¹ for the conventionally tilled field and 0.040 day⁻¹ for the non-tilled

field. The half-lives of aldicarb sulfoxide calculated from these rate constants were 15 and 16 days for the conventionally tilled and the non-tilled fields, respectively.

These half-lives are about seven times shorter than the half-lives computed from the base-catalyzed distilled water hydrolysis equation presented by Lightfoot and others (1987) for a pH of 6.5 and a temperature of 303 kelvin. Surface-catalyzed hydrolysis can explain the shorter half-lives computed from our field data. Bank and Tyrrell (1984), Bromilow and others (1986), and Lightfoot and others (1987) have shown that distilled water hydrolysis rates are considerably lower than those measured where a solid phase is present.

Smith and Parrish (1993) reported half-lives of 13 to 20 days for total aldicarb residues in a sandy loam soil. In a study conducted in sandy loam soils in the Western United States, half-lives of 15 to 65 days for total aldicarb residues were reported (Jones, 1987). Hornsby and others (1990) showed that the half-life of aldicarb metabolites taken collectively averaged about 69 days for a sandy soil in Florida. In general, the half-lives reported in these studies were somewhat higher than those computed in this study. However, the effects of surface-catalyzed hydrolysis on aldicarb sulfoxide degradation are somewhat greater in the clay-rich Falaya silt loam soils of the sampling plots in this study.

CONCLUSIONS

The aldicarb sulfoxide concentration distributions support sampling in multiple positions within transects perpendicular to the orientation of the rows. Multiple transects were proven necessary to achieve field representation for a given transect position. This sampling strategy provided suitable data sets for calculating representative and statistically defensible field values.

Base-catalyzed and surface-catalyzed hydrolysis appeared to be the most important mechanisms for the disappearance of aldicarb sulfoxide in the soil profile in both tillage systems. Degradation rate constants were 0.046 day⁻¹ for the conventionally tilled field and 0.040 day⁻¹ for the non-tilled field. The half-lives of aldicarb sulfoxide were 15 and 16 days for the conventionally tilled and the non-tilled field, respectively. Horizontal transport was negligible, and vertical transport was minimal for both tillage systems.

REFERENCES

- Bank, S. and R. J. Tyrrell, 1984. Kinetics and Mechanism of Alkaline and Acidic Hydrolysis of Aldicarb. Journal of Agricultural and Food Chemistry 32:1223-1232.
- Bromilow, R. H., G. G. Briggs, M. R. Williams, J. H. Smelt, L. G. M. Th. Tuinstra and W. A. Traag, 1986. The Role of Ferrous Ions in the Rapid Degradation of Oxamyl, Methomyl, and Aldicarb in Anaerobic Soils. Pesticide Science 17:535-547.
- Castellan, G. W., 1983. Physical Chemistry, Third Edition. The Benjamin/Cummings Publishing Company, Menlo Park, CA, 943 pp.
- Dick, W. A., R. J. Roseberg, E. L. McCoy and W. M. Edwards, 1989. Surface Hydrological Response of Soils to No-tillage. Soil Science Society of America Journal 53:1520-1526.
- Fukuto, T. R., 1987. Organophosphorus and Carbamate Ester: The Anticholinesterase Insecticides, Fate of Pesticides in the Environment: Proceedings of a Technical Seminar, pp. 5-18. Division of Agriculture and Natural Resources, University of CA, Oakland, CA, 158 pp.
- Hall, J. K., M. R. Murray and N. L. Hartwig, 1989. Herbicide Leaching and Distribution in Tilled and Untilled Soil. Journal of Environmental Quality 18:439-445.
- Hansen, J. I. and W. H. Spiegel, 1983. Hydrolysis Studies of Aldicarb, Aldicarb Sulfoxide, and Aldicarb Sulfone. Environmental Toxicology and Chemistry 2:147-153.
- Hornsby, A. G., P. S. C. Rao and R. L. Jones, 1990. Fate of Aldicarb in the Unsaturated Zone Beneath a Citrus Grove. Water Resources Research 26(10): 2287-2302.

- Isensee, A. R., R. G. Nash and C. S. Helling, 1990. Effect of Conventional Vs. No-tillage on Pesticide Leaching to Shallow Groundwater. Journal of Environmental Quality 19:434-440.
- Jones, R. L., 1987. Aldicarb Environmental Fate Studies Conducted in the Western United States. Fate of Pesticides in the Environment: Proceedings of a Technical Seminar, pp.105-108. Division of Agriculture and Natural Resources, University of CA, Oakland, CA, 158 pp.
- Jones, R. L., A. G. Hornsby and P. S. C. Rao, 1988. Degradation and Movement of Aldicarb Residues in Florida Citrus Soils. Pesticide Science 23:307-325.
- Lemely, A. T. and W. Z. Zhong, 1984. Hydrolysis of Aldicarb, Aldicarb Sulfoxide, and Aldicarb Sulfone at Parts Per Billion Levels in Aqueous Medium. Journal of Agricultural and Food Chemistry 32:714-719.
- Lightfoot, E. N., P. S. Thorne, R. L. Jones, J. L. Hansen and R. R. Romine, 1987. Laboratory Studies on Mechanisms for the Degradation of Aldicarb Sulfoxide and Aldicarb Sulfone. Environmental Toxicology and Chemistry 6:377-394.
- Metcalf, R. L. 1971. Structure-Activity Relationship for Insecticidal Carbamate. Bulletin of the World Health Organization 44:43 ff.
- Rao, P. S. C., K. S. V. Edvardsson, L. T. Ou, R. E. Jessup, P. Nkedi-Kizza and A. G. Hornsby, 1986. Spatial variability of pesticide sorption and degradation parameters, in Garner, W. Y., R. C. Honeycutt, and H. N. Nigg, Evaluation of Pesticides in Ground Water, ACS Symposium Ser. 315, American Chemical Society, Washington D.C.
- Seiber, J. N., 1987. Solubility, Partition Coefficient, and Bioconcentration Factors. Fate of Pesticides in the Environment: Proceedings of a Technical Seminar, pp.53-59. Division of Agriculture and Natural Resources, University of CA, Oakland, CA, 158 pp.
- Smith, C. N. and R. S. Parrish, 1993. A Field Study to Evaluate Leaching of Aldicarb, Metolachlor, and Bromide in a Sandy Loam Soil. Journal of Environmental Quality 22:562-577.
- Smith, C. N., R. S. Parrish and R. F. Carsel, 1987. Estimating Sample Requirements for Field Evaluation of Pesticide Leaching. Environmental Toxicology and Chemistry 6:343-357.
- U.S. Department of Agriculture, Soil Conservation Service, 1989. Soil Survey of Shelby County, Tennessee. U.S. Government Printing Office, Washington, D.C., 56 pp. and 90 maps.

RELATION BETWEEN NITRATE CONCENTRATIONS AND POTENTIAL NITRATE SOURCES FOR WATER-TABLE AQUIFERS IN THE BEAVER CREEK WATERSHED, WEST TENNESSEE: A STATISTICAL ANALYSIS

Shannon Williams' and Angel Roman-Mas²

ABSTRACT: Non-parametric statistical methods were used to examine the relation between nitrate concentrations and potential sources of nitrate for the water-table aquifers in the Beaver Creek watershed. Water-quality data collected in 1992 from 81 domestic wells were used for these analyses. The data showed that nitrate concentrations (measured as nitrate) exceeded 13 milligrams per liter in 16 percent of the water samples analyzed. The Spearman Rank correlation analysis indicated that nitrate concentrations are well-depth dependent. The Kruskal-Wallis test indicated that the variance in nitrate concentration can be explained in terms of selected potential nitrate-source categories. The Student-Newman-Keuls multiple comparison of the means test showed that: (1) the mean nitrate concentration for wells in the deeper than 45-meter category was lower and statistically different than the mean concentrations for the other categories, (2) the mean nitrate concentrations for categories that include septic tanks were higher and statistically different than the mean concentration for the crop category, and (3) the mean nitrate concentration for the confined animals category was higher and statistically different than the mean concentration for the crop category.

KEY TERMS: Water-table aquifers; nitrate; rural areas; statistical analyses.

INTRODUCTION ...

The increasing incidence of ground-water contamination has resulted in a need for: (1) ground-water quality and land-use data and (2) analytical tools to establish relations between anthropogenic activities and ground-water quality. Row-crop agriculture has been recognized as a potential non-point source of ground-water contamination. Water draining cropland can contribute large amounts of nitrate and other contaminants to receiving aquifers, thus impairing their suitability for designated uses. However, row crops are not grown isolated from other nitrate sources, such as septic tanks and facilities with confined animals. Accordingly, analytical tools must be able to differentiate the various nutrient sources.

Parametric and non-parametric statistical analyses are commonly used to relate anthropogenic activities to ground-water quality conditions. Although statistical analyses can provide significant insight, investigators should exercise caution in the selection of the proper statistical methods. Objectives of the evaluation and the statistical characteristics of the water-quality data and other relevant

^{&#}x27;Graduate Student, Department of Geography, University of Memphis, Memphis, TN 38152

²Hydrologist, U.S. Geological Survey, 7777 Walnut Grove Rd. Suite LLB-2, Memphis, TN 38120

variables, such as well depth and distance-to-source, also should be considered when selecting appropriate statistical methods. In performing the statistical analyses, it is assumed that each well or spring sampled and the associated water-quality data can be categorized according to potential sources of a contaminant. The categorization process should be consistent with the physiographic and hydrogeologic setting of the area.

Agriculture is vital to the economy of the Beaver Creek watershed in West Tennessee. About 70 percent of the watershed is under crop production that consists mainly of soybean, cotton, and corn. As in many rural areas in the United States, residents in the Beaver Creek watershed rely on domestic wells as their primary source of water. The residents also share a growing concern that agricultural activities could be affecting the quality of the ground water in the watershed.

In the summer of 1992, a water-quality reconnaissance of the water-table aquifers in Shelby, Tipton, Fayette, and Haywood Counties, Tennessee, was conducted by the U.S. Geological Survey, in cooperation with the Tennessee Department of Agriculture, and the University of Tennessee Agricultural Extension Service. The reconnaissance included the collection of water samples from 81 domestic wells completed in the water-table aquifers in the Beaver Creek watershed (Figure 1) (Fielder and others, 1994).

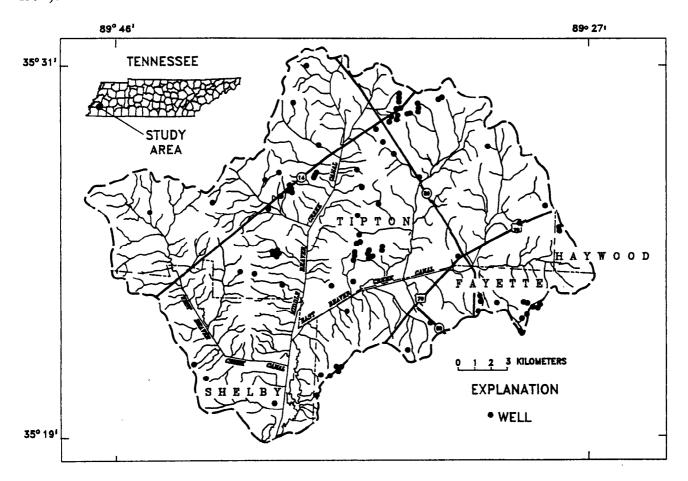


Figure 1. Location of Beaver Creek study area and sampled wells.

Nitrate concentrations (measured as nitrate) exceeding 13 mg/L (milligrams per liter) were observed in 16 percent of the water samples for wells in the Beaver Creek watershed. Concentrations above 13 mg/L can indicate influence of anthropogenic activities (Madison and Brunett, 1985). Water samples from two of the wells exceeded the primary drinking water standard for nitrate (measured as nitrate) of 45 mg/L adopted by the Tennessee Department of Environment and Conservation (1993). Ingestion

of water with high nitrate content has been found to cause methemoglobinemia in infants (Klaassen and Rozman, 1991). Statistical analyses are used in this paper to relate nitrate concentrations to potential nitrate sources in the Beaver Creek watershed.

HYDROGEOLOGY

Alluvium and fluvial deposits of Quaternary and Tertiary(?) ages make up the shallow aquifers in the Beaver Creek watershed (Parks and Carmichael, 1990a, 1990b; Parks, 1990). Alluvium, consisting of sand, gravel, silt, and clay, is present beneath the bottomland areas. Fluvial deposits, consisting of sand, gravel, minor clay, and ferruginous sandstone, are present beneath the loess in upland and valley slope areas. Locally, the alluvium and fluvial deposits are underlain by the Cockfield Formation, Cook Mountain Formation, or the Memphis Sand (Parks and Carmichael, 1990a, 1990b). The Cockfield Formation and Memphis Sand consist of sand, silt, clay, and lignite and make up the Cockfield and Memphis aquifers. The Cook Mountain Formation consists primarily of clay and is the confining unit overlying the Memphis aquifer. Hydrogeologic information for the deeper wells sampled during this investigation was insufficient to determine whether or not some of these wells are screened in the Cockfield or Memphis aquifers.

METHODS

Protocols for well selection and for sample collection and analysis have been described by Fielder and others (1994). A detailed land-use inventory was conducted for each well included in the sampling. Land use near each well site was described and distances and topographic gradients between wells and potential nitrate sources were recorded. Well construction data were provided by well owners. Well records from the Tennessee Department of Environment and Conservation (TDEC) were used to verify information obtained from well owners.

The selection of statistical methods used in this paper follows the discussion by Harris and others (1987) and Montgomery and others (1987). All statistical analyses were performed using CoStat statistical software (CoHort, 1990) (Any use of trade, product, or firm names in this article is for descriptive purposes only and does not imply endorsement by the U.S. Government).

RESULTS AND DISCUSSION

Well Depth and Distance-to-Source

Researchers have shown that the nitrate concentration in ground water is a function of well depth and distance-to-source (Madison and Brunett, 1985; Murphy, 1992). Correlation analyses can be conducted to assess the concentration/well depth and the concentration/distance-to-source dependencies. The well depth and distance-to-source at which the water quality of the well is no longer affected by a particular source can be determined from these analyses. These outer limits should be integrated into the classification process. If nitrate concentrations are dependent on well depth and distance-to-source, then depths and distances among potential nitrate-source categories cannot be statistically different for a comparison among categories to be valid.

Well depth and nitrate concentration data were not normally distributed; they were skewed to the right (positive skewness). Accordingly, a non-parametric correlation test (Spearman Rank) was used. Nitrate concentrations were observed to decrease with depth and the Spearman Rank coefficient was -0.55 (Figure 2). This analysis indicates that nitrate concentrations are well-depth dependent; therefore, well depth must be considered when evaluating the relation between potential nitrate sources and nitrate concentrations.

The y-intercept resulting from the correlation between well depth and nitrate concentration (45 m)

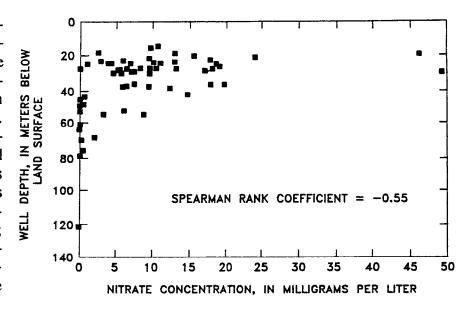


Figure 2. Relation between nitrate-as-nitrate concentration and depth of sampled wells.

was used to segregate the data into two categories (less than 45 m and greater than 45 m in depth). The segregated data were subjected to a series of statistical analyses to prove or disprove the null hypothesis: the variance in nitrate concentration between the two well-depth categories is random. Nitrate concentrations for both well-depth categories also showed positive skewness. Therefore, the Mann-Whitney test (non-parametric) was utilized to test the null hypothesis. The test indicated that the variation in the nitrate concentration has a low probability (10%) of being due to random variation (Figure 3). Therefore, the variance in the nitrate concentration can be explained in terms of the selected well-depth categories. These analyses indicate that nitrate concentration for wells deeper than 45 m are not affected by anthropogenic activities.

Distance-to-source (nearest source) and nitrate-concentration data were also positively skewed. Because these data were not normally distributed, the Spearman Rank non-parametric correlation test

was used. A significant relation between the two variables was not observed; the Spearman Rank coefficient was -0.21 (Figure 4).

Nitrate-Source Categories

In order to apply statistical analyses to relate the nitrate concentrations to potential nitrate sources, each well sampled was classified into one potential nitrate-source category. Each category included one or more nitrate sources. Only those sources that were relevant to the individual well were considered. The

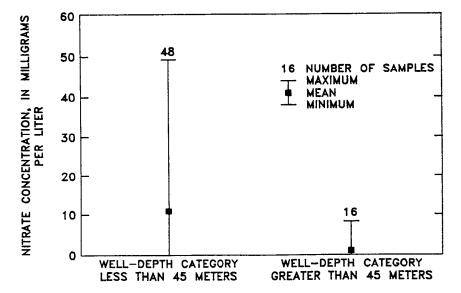


Figure 3. Summary of nitrate-as-nitrate concentration data by well-depth category.

hydrogeologic characteristics of the aquifer and well hydraulics were used for determining which were the potential contributing sources. However, the available hydrogeologic and hydraulic data were somewhat limited.

Distances and topographic gradients were used to classify the sampled wells into potential nitrate-source categories. The direction of ground-water flow in the shallow aquifers was assumed to generally correspond to topographic gradients. Accordingly, all potential nitrate sources upgradient were

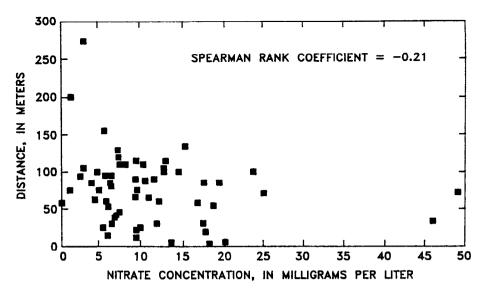


Figure 4. Relation between nitrate-as-nitrate concentration and distance to nearest potential nitrate source for sampled wells.

considered in the classification. The sampled wells are low-yield domestic wells that are pumped intermittently to holding tanks. Pumping periods are usually less than 5 minutes and pumping rates are less than 38 liters per minute. The well radius of influence is assumed to be relatively small. Therefore, potential nitrate sources more than 15 m downgradient were not considered in the classification.

Potential nitrate sources considered in the classification were: (1) cropland (fertilizer application), (2) septic tank systems, and (3) confined animal facilities. These sources resulted in six combinations of categories (Table 1). The wells deeper than 45 m were considered as an individual category. Nine of the wells sampled did not fall into any of the selected categories and were not used for further analyses.

Statistical Analyses

TABLE 1. Classification of wells by potential nitrate sources

Potential Nitrate Sources	No. of Wells
Septic tanks only (ST)	12
Septic tanks and crops (ST/CR)	26
Septic tanks and confined animals (ST/CA)	9
Crops only (CR)	3
Confined animals only (CA)	3
Crops and confined animals (CR/CA)	3
Wells more than 45 meters deep (TD) Wells that did not fit into any of the	16
above categories	9
TOTAL	81

In order to evaluate the relative contribution of potential

nitrate sources to nitrate concentrations, the variance in the nitrate concentrations should be explained in terms of the selected potential nitrate-source categories. The Kruskal-Wallis test (non-parametric) was used for this evaluation. Skewness tests indicated that nitrate concentration data for several of the categories were not normally distributed; therefore, parametric tests could not be used. The Kruskal-Wallis test determines only the probability at which the variance for a particular variable (nitrate concentration) is due to random variation (null hypothesis). If the null hypothesis is rejected, the

selected data treatments (potential nitrate-source categories) can be a plausible explanation for the observed variance. Results of the Kruskal-Wallis test indicated that the variance in the nitrate concentration has a low probability (10°) of being due to random variation. Therefore, the variance in nitrate concentration is explained in terms of the selected potential nitrate-source categories.

Because more than two categories were considered, the analysis of the variances was followed by a multiple comparison of the means. Student-Newman-Keuls multiple comparison test (nonparametric) was used to rank nitrate-concentration means and to determine which means were statistically different at a 0.05 level of significance (Figure 5). showed that: (1) the tests mean concentration for wells classified as deeper than 45 m was lower and statistically different than the means for the other six categories, (2) the means for categories that in-

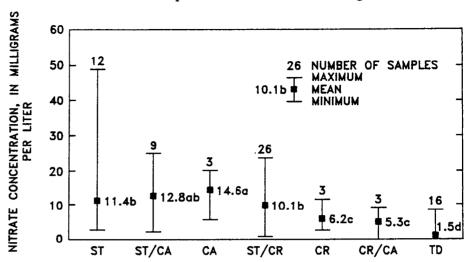


Figure 5. Summary of nitrate-as-nitrate concentration data for selected potential nitrate-source categories (letter following mean value represents populations that are not statistically different at the 0.05 probability as determined by the Student-Newman-Keuls multiple com-parison test. ST, septic tank; CA, confined animals; CR, crops; TD, wells deeper than 45 meters).

clude septic tanks were higher and statistically different than the mean for the crop category, and (3) the mean for the confined animals category was higher and statistically different than the mean for the crop category.

The test of the variance and the multiple comparison of the means test assumed that the nitrate data were stationary in time and space. These assumptions were supported by: (1) the relatively short time period (July-August) in which the samples were collected, (2) the uniformity in the climatic conditions during the sampling period (dry season), (3) the absence of on-going agricultural activities that could affect nitrate concentrations (planting season ends in early June), and (4) the relatively low hydraulic conductivity of these aquifers. For these reasons, the data were assumed to be independent and serial correlations were not conducted.

Because nitrate concentrations were determined to be dependent on well depth, the Kruskal-Wallis test was used to compare well-depth distribution among the various potential nitrate-source categories. The results showed that the variation in well depth among the various categories can be explained by random variation (p = 0.35) (Figure 6). Therefore, well depths do not bias the comparison among the nitrate-source categories.

CONCLUSIONS

Results from the Spearman Rank correlation test indicated that nitrate concentrations are well-depth dependent. Therefore, well depth had to be considered when evaluating nitrate source-concentration relations. The results of the Mann-Whitney test supported the assumption that nitrate concentrations for wells deeper than 45 m are not affected by anthropogenic activities.

The Kruskal-Wallis test indicated that the variance in nitrate concentration can explained in terms of the selected nitrate-source catepotential gories. The Student-Newman-Keuls multiple comparison of the means test showed that: (1) the mean nitrate concentration for wells in the deeper than 45 m category was lower and statistically different than the means for the other six categories, (2) the mean nitrate concentrations for categories that include septic tanks were higher and statisti-

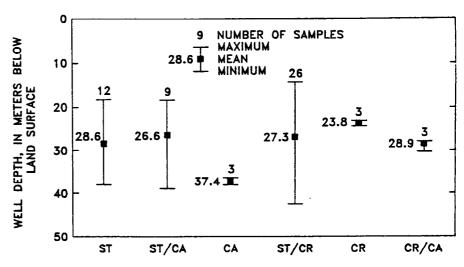


Figure 6. Relation between well depth and selected potential nitrate-source categories (ST, septic tank; CA, confined animals; CR, crops).

cally different than the mean for the crop category, and (3) the mean nitrate concentration for the confined animals category was higher and statistically different than the mean for the crop category.

The results of these analyses are limited by the relatively small sample sizes for some of the potential nitrate-source categories. However, the number of wells per unit area for a given category is relatively large when compared to similar investigations reported in the literature. Furthermore, the distribution of sampled wells among the various categories reflects the occurrence of potential nitrate sources with respect to well location throughout the watershed.

REFERENCES

CoHort Software, 1990. CoStat statistical software manual revision 4.20. CoHort Software, Berkeley, California, 278 p.

Fielder, A.M., A. Roman-Mas and M.W. Bennett, 1994. Reconnaissance of Ground-Water Quality at Selected Wells in the Beaver Creek Watershed, Shelby, Fayette, Tipton, and Haywood Counties, West Tennessee, July and August 1992. U.S. Geological Survey Open-File Report 93-366, 28 p.

Harris, J., J.C. Loftis and R.H. Montgomery, 1987. Statistical Methods for Characterizing Ground-Water Quality. Ground Water 25(2):185-193.

Klaassen, C.D. and K. Rozman, 1991. Absorption, Distribution, and Excretion of Toxicants in Amdoor, M.O., J. Doull and C.D. Klaassen, eds. Toxicology: The Basic Science of Poisons. Pergamon Press, Elmsford, NY, chapter 3, pp. 50-85.

Madison, R.J. and J.O. Brunett, 1985. Overview of the Occurrence of Nitrate in Ground Water of the United States. U.S. Geological Survey Water-Supply Paper 2275, pp. 93-105.

Montgomery, R.H., J.C. Loftis and J. Harris, 1987. Statistical Characteristics of Ground-Water Quality Variables. Ground Water 25(2):176-184.

Murphy, E.A., 1992. Nitrate in Drinking Water Wells in Burlington and Mercer Counties, New Jersey. Journal of Soil and Water Conservation 47(2):183-187.

Parks, W.S., 1990. Hydrogeology and Preliminary Assessment of the Potential for Contamination of the Memphis Aquifer in the Memphis Area, Tennessee. U.S. Geological Survey Water-Resources Investigations Report 90-4092, 39 p.

- Parks, W.S. and J.K. Carmichael, 1990a. Geology and Ground-Water Resources of the Cockfield Formation in Western Tennessee. U.S. Geological Survey Water-Resources Investigations Report 88-4181, 17 p.
- Parks, W.S. and J.K. Carmichael, 1990b. Geology and Ground-Water Resources of the Memphis Sand in Western Tennessee. U.S. Geological Survey Water-Resources Investigations Report 88-4182, 30 p.
- Shapiro, S.S., M.B. Wilk and H.J. Chen, 1968. A Comparative Study of Various Tests for Normality. American Statistical Association Journal, pp.1343-1372.
- Tennessee Department of Environment and Conservation, 1993. Regulations for Public Water Systems and Drinking Water Quality. Tennessee Department of Environment and Conservation, Division of Water Supply, chapter 1200-5-1, 174 p.

FATE AND TRANSPORT OF NITROGEN IN AN AGRICULTURAL WATERSHED

A. Roman-Mas¹, H.H. Cochrane², J.A. Smink³, and S.J. Klaine³

The fate and transport of nitrogen in an agricultural watershed in West Tennessee was studied at the field and watershed levels. Runoff and water-quality data for four ditches draining row crop fields in the Beaver Creek watershed were used to quantify the amount of nitrogen exported from cropland. Drainage areas for these fields ranged from 7 to 200 hectares, with about 75 to 95 percent of the drainage areas under row crop cultivation. Runoff and water-quality data for a 3rd-order and a 5th-order stream site downstream of the ditches in the Middle Beaver Creek Canal were used to evaluate potential changes in nitrogen speciation and load in the watershed streams. Drainage areas for these sites were 1,200 and 12,000 hectares, respectively, with about 70 percent of the areas under row crop cultivation.

Data from about 100 rainfall-runoff events were used for this study. Discharge records were developed from modified U.S. Geological Survey operational procedures. Discrete water samples were collected at intervals ranging from 5 to 30 minutes and analyzed for total and dissolved nitrogen species.

Data for the four ditches show that nitrogen in runoff from row crop fields was dominated by the suspended-organic fraction. Crop residue in different stages of decomposition accounted for about 80 percent of the total nitrogen export. Nitrite plus nitrate accounted for about 20 percent of the total nitrogen export from these fields. Ammonia was detected in trace concentrations only. Some seasonal patterns were evident. Late winter and early spring rainfall-runoff events accounted for 75 percent of the suspended-organic nitrogen exported annually. Late spring rainfall-runoff events accounted for 80 percent of the nitrite plus nitrate exported annually. Growing-season rainfall-runoff events accounted for less than 10 percent of the annual nitrite plus nitrate export.

Data for the 3rd- and 5th- order stream sites show that no significant changes in the nitrogen chemical speciation or the load per unit area occurred downstream from the row crop fields. Data for these sites show that channelized streams in this watershed are efficient at conveying runoff as well as dissolved and particulate nitrogen in runoff. However, these streams do not change the nitrogen content of cropland runoff.

^{&#}x27;Hydrologist, U.S. Geological Survey, 7777 Walnut Grove Rd., Suite LLB2, Memphis, Tennessee 38120.

²Graduate Student, University of Memphis, Department of Geography, Memphis, Tennessee 38152. ³Graduate Student and Associate Professor, respectively, Department of Environmental Toxicology and The Institute for Wildlife and Environmental Toxicology, Clemson University, Pendleton, South Carolina 29670.

SUSPENDED-SEDIMENT YIELD AND CHANNEL EVOLUTION IN A SMALL AGRICULTURAL CATCHMENT

Robert W. Stogner, Sr. 1 and Timothy H. Diehl2

Research on suspended-sediment yield from agricultural land and channel evolution in permanent field ditches is being conducted in a small catchment in the Beaver Creek watershed of West Tennessee. This catchment has an area of about 11 hectares, 90 percent of which has been planted in cotton annually since 1989. The soils are Grenada, Loring, Memphis, and Adler silt loams.

Surface-water discharge was measured from July 9, 1991 to October 15, 1993, except for short periods of equipment failure. Rainfall during this period was typical of average conditions. Total runoff volume during 95 measured storms was 61,000 cubic meters. The median elapsed time from initial hydrograph rise to hydrograph peak for 20 selected storms was 15 minutes.

Suspended-sediment concentration was measured during 43 storms, typically at a sampling interval of 5 minutes. The total runoff volume for these storms was 30,000 cubic meters, about 49 percent of the total runoff volume during the study period. The measured total suspended-sediment yield from these storms was 170 metric tons (mt). Suspended-sediment yield from storms during which sediment concentration was not measured was estimated to be about 200 mt using the regression of storm load on storm volume. The 90-percent confidence interval for this estimate, based on the standard error of the regression, is from 100 to 460 mt. The estimated total suspended-sediment yield from all 95 storms was 370 mt, with a 90-percent confidence interval from 250 to 640 mt.

Evolution of the permanent ditch draining the cotton field was studied. The ditch is about 22 m long, about 1 m wide at the bank tops, and from 0.6 to 0.9 m deep. Its bed slope is about 1 percent. During the study period, the thalweg of the ditch degraded an average of 0.11 m, and a net volume of 5 cubic meters of bed and bank material was eroded, contributing an estimated 7 mt of suspended sediment. Erosion in the ditch contributed between 1 and 3 percent of the total suspended-sediment yield from the catchment.

¹Hydrologist, U.S. Geological Survey, 7777 Walnut Grove Blvd., Suite LLB2, Memphis, Tennessee 38120.

²Hydrologist, U.S. Geological Survey, 810 Broadway, Suite 500, Nashville, Tennessee 37203.

MECHANISTIC EVALUATION OF PESTICIDE TEMPORAL PATTERNS FOR A FIRST-ORDER STREAM

Angel Roman-Mas¹, Carol P. Weisskopf², and Stephen J. Klaine²

Processes and factors affecting the temporal patterns in the concentration of aldicarb and its metabolites were evaluated for a first-order stream in the Beaver Creek watershed, West Tennessee. Mathematical algorithms proposed to explain solute transport mechanics for soil-applied chemicals and, in particular, the mass transfer of a chemical from the soil solution into runoff were examined.

Two general patterns in the concentration distribution of aldicarb and its metabolites were observed during rainfall-runoff events. In the early stages of the event, the natural log of the concentration increased linearly with time-and inversely in proportion to the rainfall-runoff ratio. The duration of this initial pattern was similar to the basin response (lag) time. The pattern ended when the concentration peaked. After peak concentration, a second pattern was observed. The natural log of the concentration decreased exponentially with time and was proportional to the rainfall-runoff ratio. These patterns in the concentration of aldicarb and its metabolites are consistent with the rate-limited transfer algorithm used in physically based models and are inconsistent with the instantaneous-equilibrium algorithm used in lumped-parameter models.

Peak concentrations for aldicarb and its first metabolite, aldicarb sulfoxide, for rainfall-runoff events decreased exponentially with elapsed time between application and an event. The rate at which the peak concentrations of aldicarb and aldicarb sulfoxide in runoff decreased with time was similar to the degradation rates in the soil profile. The peak concentrations of aldicarb sulfone, the last metabolite in the oxidation of aldicarb, increased exponentially with time until day 18 and decreased exponentially with time after that. These patterns were consistent with those observed in soils. Multiple regression analyses indicated that peak concentration in runoff for a given rainfall-runoff event is controlled primarily by the concentrations in the soil solution.

^{&#}x27;Hydrologist, U.S. Geological Survey, 7777 Walnut Grove Rd., Memphis, Tennessee 38120.

²Assistant Professor and Associate Professor, respectively, The Institute for Wildlife and Environmental Toxicology and Department of Environmental Toxicology, Clemson University, Pendleton, South Carolina 29670.

RESEARCH AND MONITORING NEEDS FOR THE IMPLEMENTATION OF THE USDA WATER QUALITY HUA PROGRAM: THE BEAVER CREEK WATERSHED PROJECT

Howard C. Hankin¹ and George F. Smith²

Abstract: The hydrologic unit area (HUA) program was initiated because of increasing concern for water resources in rural United The HUA program is a major element of the President's Water Quality Initiative within the U.S. Department of Agriculture. Previous experiences with rural water-quality programs have demonstrated the need for intensive monitoring programs to identify the sources and magnitudes of agricultural non-point source and to document water-quality improvements. pollutants Additionally, many questions remain about the effects of best management practices on the transport and fate of potential contaminants, including sediment, nutrients and pesticides. HUA projects should be designed to address these issues and concerns. They should also serve as demonstration projects to increase awareness in water-quality issues within the agricultural community.

KEY TERMS: agriculture; nonpoint source pollution; water quality.

INTRODUCTION

Concern for water-quality issues in rural United States has increased in the past decade. Nonpoint source pollution and watershed protection have become areas of special attention (Knopman and Smith, 1993). As a result of this concern, the President's Water Quality Initiative has initiated the hydrologic unit area (HUA) program within the U.S. Department of Agriculture (USDA). The HUA program targeted watersheds at locations throughout the United States where water quality was considered impaired as a result of agricultural non-point source pollution. The primary objective of the HUA program was to improve water quality in these watersheds through the implementation of best management practices (BMPs).

The Soil Conservation Service (SCS), Agricultural Extension Service (AES) and Agricultural Stabilization and Conservation Service (ASCS) are USDA agencies that each play a major role in the HUA program. The SCS, AES and ASCS have the responsibility to provide technical, educational and financial assistance, respectively, to farmers. Landowners can choose among many

¹Resource Conservationist, USDA Soil Conservation Service, 675 U.S. Courthouse and Federal Building, 801 Broadway, Nashville, TN 37203. ²Professor, Resource Development, TN Agricultural Extension Service, University of Tennessee, PO Box 1071, Knoxville, TN 37901.

alternative resource management options with different production, economic and environmental consequences. The USDA representatives must be able to offer landowners the most accurate information available to assist them in making the best possible decisions for each unique situation. This can only be accomplished with a reliable understanding of the targeted watershed.

Experiences from the Rural Clean Water Program and agriculture nonpoint source studies have demonstrated the need for intensive monitoring programs to achieve reliable information about the watershed (Richards and Baker, 1993). Successful monitoring will help identify the sources and magnitudes of agricultural non-point source pollution (NPS). Continued water-quality monitoring was also deemed important to document improvements in water quality resulting from changes in land-management practices. Additionally, many questions remain about the effects of BMPs on the transport and fate of potential contaminants, including sediment, nutrients and pesticides (Cooper, 1993).

Another objective of the HUA program is to demonstrate the effectiveness of voluntary programs at mitigating water quality in the targeted watersheds. This feedback is vital to farmers, policy makers and the general public. Demonstrating improved water quality as a result of BMPs encourages other farmers to participate and become involved. The HUA projects should demonstrate economic feasibility, and improve environmental sensitivity and public acceptability. These are key determinants in the decision-making process of contemporary agriculture.

Successful implementation of many HUA projects has been hindered by the absence of data and modeling capabilities that (1) accurately define the nature and extent of the impacts of agricultural activities on water quality, and (2) support the selection and implementation of BMPs that best fit conditions at the field and at the watershed level. The intent of this paper is to focus on several major needs USDA agencies have in order to better assist landowners in managing their natural resources. Examples will be drawn from the Beaver Creek HUA in Southwestern Tennessee to highlight successes and future needs.

Beaver Creek HUA Project

The Beaver Creek HUA project in Southwestern Tennessee was initiated in 1990 as a result of farmers' concern about NPS pollution. Reasons for the concern ranged from statements in the national media attributing significant water-quality impairments to modern agriculture, to an assessment level evaluation conducted by state agencies in Tennessee. The water-quality degradation in the Beaver Creek Watershed (BCW) was attributed to agricultural activities (Thompson, 1993). A comprehensive water-quality monitoring program was initiated in addition to implementing BMPs in the Beaver Creek HUA to achieve the objectives of the HUA program.

RESEARCH NEEDS

Research activity that would improve the ability to manage the natural resource base is needed. The goals of this research effort should include: (1) improving our ability to determine the nature and extent to which agricultural activities impact water quality and threaten environmental integrity, (2) improving our ability to differentiate agricultural NPS pollution from other sources of pollution and from natural conditions, (3) developing and evaluating monitoring and sampling strategies that accurately represent water quality, (4) understanding the processes and factors that control the transport and fate of agricultural pollutants, (5) assessing the validity of present criteria used to set standards for water quality, (6) calibrating and validating water-quality models.

The effects of agricultural BMPs such as conservation tillage, nutrient management and pesticide management have been studied throughout the country (Sharpley and others, 1988; Jayachandron and others, 1993). Inconsistent results have been obtained by virtue of a number of site specific factors influencing the processes of soil erosion and nutrient pesticide movement (Betson and Marius, 1969; Griffin, 1992)). More research is needed to quantify site specific variables which determine the effectiveness of BMPs used alone and in various combinations.

The use of the simulation model, CREAMS, in Pennsylvania, resulted in substantial variance among sites and practices with regards to effectiveness of BMPs and nutrient management programs (NMPs) in reducing off-site sediment and nutrient loads (Hamlett and Epp, 1994). It was concluded that "typically there is not a single BMP or NMP that is adequate for controlling pollutant losses from agricultural lands. Therefore, to control sediment and nutrient losses from agricultural areas, control programs should emphasize the use of improved NMPs with the more traditional BMPs", (Hamlett and Epp, 1994). It was also concluded that field conditions such as slope, crop rotation, and the predominant transport pathways should be considered when selecting BMPs and NMPs.

A study using the paired watershed approach in the limestone soils of northern Alabama was conducted by Soileau and others (1994). Analyses of the data collected over a 6-year period suggested a strong effect of tillage on runoff water. Specifically, a greater volume of runoff water occurred with the use of conservation tillage than conventional tillage. However, soil losses for both tillage systems were below soil tolerance (T), but significantly lower for conservation tillage when soil loss calculations were determined using the erosion index. Also, the presence of a grass filter strip together with crop residue cover in the winter and spring also helped minimize sediment loss from the watershed, especially with conventional tillage.

Soileau and others (1994) found that tillage operations appeared to have no significant effect on nitrogen (N) losses, either as Kjeldahl nitrogen or as NO₃-N. Phosphorus (P) runoff tended to be greater with conservation tillage than with conventional tillage. This contradicts studies done by Sharpley

and others (1991) who found a reduction in nutrient (N and P) transport from a sorghum field located in the southern plains with conservation tillage. The dissimilar results could be explained in part by the unique site variables. Most of the annual P runoff from the Alabama site occurred with the filtered water component instead of the sediment-bound fraction because this site had a relatively large volume of runoff water and small sediment loss. Another explanation could be the use of different sampling and analytical techniques.

Again, these different research projects demonstrate the variability in results, possibly due to sampling or research strategies, geographical location, soil types, climate or even the duration of the project. They also demonstrate the need for additional research utilizing consistent sampling methodology so that legitimate comparisons between data sets become possible.

An important factor that must be considered is the financial feasibility of the BMP to the farmer. As concluded by Hamlett and Epps (1994), various approaches may prove effective in the field, but they may negatively influence the farm enterprise or may not be financially feasible in a real-world situation. Therefore, economic analyses of all practices and programs are needed.

USDA agency personnel need to understand how the implementation of no-till and combinations of other BMPs behave with respect to the fate of nutrients and pesticides. This includes the migration of agri-chemicals through different types of soils. The agent must be able to integrate that knowledge with the myriad combinations of crops, vegetative cover, soil types, field slopes, climate and economic feasibility. Ultimately a system of conservation and nutrient practices which will provide beneficial effects to the environment must be achieved.

Additional information is needed on the types of vegetation best suited for erosion control and nutrient and/or pesticide uptake and deposition under the varying climatic and geographic conditions. An enormous amount of research has been conducted on these topics, but the results need to be integrated to provide the best recommendations for land-users.

The success of BMPs is dependant upon the types of vegetation associated with these practices. Promoting certain types of vegetation as winter cover crops, conservation cropping systems or in filter strips and vegetated waterways has proven to be extremely successful at reducing the transport of soil, nutrients and pesticides that have an affinity to clay. For example, it has been shown that grasses and legumes were effective for erosion control purposes in row crops. It has also been demonstrated that vegetative barriers (known as stiff grass hedges) can reduce sediment yield and decrease runoff velocity (Dabney and others, 1993).

The introduction of legumes into a conservation cropping system has proven to be useful both as erosion control and adding nitrogen to the soil for the following seasons crop. It has been demonstrated that some winter cover crops, such as rye, can scavenge nitrogen from previous applications and prevent its movement through the soil profile into ground water. However, some of these same cover crops may actually promote NPS pollution into

adjacent water systems. An example of this condition has been shown with P leaching from cotton foliage, wheat straw (Soileau and others, 1994), and the canopy of sorghum and soybeans (Sharpley, 1981). The P leached from the crops was a major portion of the total P runoff in these studies. This P leached from plant tissue, may partly explain why cotton plus rye in the conservation tillage cropping system had more total P runoff loss than cotton only in the conventionally tilled system in the northern Alabama project (Soileau and others, 1994).

Recognizing the sources of nutrient loading is a critical factor in developing natural resource plans. All of the pollution sources potentially affecting soil, water, air, plants, animals, and humans must be identified as plans are developed to alleviate problems affecting designated use impairments. If potential nonpoint pollution sources are not accounted for, the restoration plan may never achieve its intended goals. Thus, we must continue to develop our understanding of both the natural and human influences on the environment.

Monitoring Needs

Optimized sampling strategies to assess agricultural NPS pollution and BMPs must be considered as the foundation for determining baseline conditions, sources of contaminants, and the effectiveness of decisions made and actions taken. Meeting these needs requires an intensive monitoring program at both the field, sub-watershed, and large watershed level (Richards and Baker, 1993). Analyses and subsequent conclusions are only as reliable as the methodology that is utilized. Therefore, it is imperative that quality assurance and quality controls are employed as part of the monitoring strategy.

There are additional concerns when monitoring nonpoint pollution parameters in large watersheds. These concerns include site selection and multiple nonpoint pollution sources. Proper selection of sampling sites is imperative when sampling at the watershed level. One should be aware of hydrologic factors, such as backwater at the mouth of a watershed, that could influence the chemical and biological parameters being assessed. Caution must be taken, as these conditions generally yield higher concentrations of the contaminants being evaluated owing to the retardation of water flow.

Another concern that a successful monitoring program must address is that of multiple nonpoint pollution sources. It is extremely difficult, if not impossible, to differentiate sources of NPS pollution at the watershed level. A thorough reconnaissance of the watershed must be done to catalog the possible sources of pollutants. Specific sites should be monitored to determine the NPS contribution of an area to the watershed. Additional sampling should be done downstream to determine if the NPS pollution has been removed from the system (by assimilation and breakdown) or has migrated downstream. Without characterizing this process, it is possible to incorrectly identify and target a source as the primary contributor.

Many of the desired water-quality goals were developed for low-flow conditions. However, NPS pollutant loading occurs primarily during rain events. These elevated levels of NPS pollutants last for relatively short periods of time. This implies that chemical monitoring must be intensive during the rain event to accurately characterize the NPS pollutant load, especially in small basins (Richardson and Baker, 1993). Temporal and spatial considerations are critical to a successful chemical monitoring program. A great deal of financial and personnel resources are required to accurately characterize the NPS pollution in a watershed.

Many agencies are turning to biological monitoring to avoid the problems associated with a chemical monitoring program. However, there are concerns with the use of biological monitoring to draw conclusions regarding NPS pollutants. For example, the biological database is presently too small to establish scientifically defensible limits for specific geophysical regions (Griffin and others, 1992). Preliminary research must be done to develop an acceptable baseline data set. This is essential to factor out the inherent diversity of the watershed system, as well as seasonal and annual fluctuations. Acceptable ranges must be established to account for natural variability.

Many sampling protocols are available for biological monitoring (Byl and Roman-Mas, 1994). However, these protocols were developed for specific hydrologic and geographic conditions that may or may not apply to the watershed under investigation. For example, the use of the sensitive taxa (Ephemeroptera, Trichoptera, and Plecoptera) to evaluate water quality may not be applicable to watersheds that do not supply the correct habitat for these organisms. It is entirely possible that these organisms never inhabited the watershed in question, and therefore, accurate comparisons cannot be made.

Despite the many needs of monitoring programs, accurate biological and chemical assessments are necessary in order to properly characterize the contaminants and their sources. Otherwise, application of conservation management systems, or BMPs, to restore or enhance the impaired water quality will be ineffective.

The Beaver Creek HUA Program

The Beaver Creek HUA project has provided a vehicle to conduct intensive monitoring and research in real farm situations. It has presented the opportunity to improve methodologies for surface and ground-water monitoring. Experience gained from several years of pre-BMP monitoring has allowed the Beaver Creek program to develop a considerable data base to use in water-quality models. The intensive studies conducted in the Beaver Creek HUA have also improved our understanding of in-stream assimilation processes. This has led to the development and testing of innovative new BMPs, such as a constructed wetland to mitigate runoff from a field of row crops.

The U.S. Geological Survey, in cooperation with USDA, local and state agencies, is conducting a comprehensive assessment of the effect of agricultural activities on water quality and the effectiveness of BMPs in the Beaver Creek Watershed. The main goal of this project is to develop tools that can be used by federal and state agencies in their resource management programs. A holistic watershed approach has been adopted for this program. Intensive water sampling has been done during rain-events to get chemical data and characterize the transport of NPS pollution in the watershed. The chemical monitoring has examined both surface and ground water. Biological samples have also been gathered in connection with the chemical and geomorphology studies. Some of the results of these monitoring activities can be found in these proceedings.

SUMMARY

Effectively addressing NPS water-quality problems from agricultural sources depends upon (1) careful identification of the NPS problems and their source, (2) implementation of effective and cost efficient practices to address the problems identified, and (3) the capability to demonstrate the benefits of resource management decisions on water quality. Research is needed to aid in identifying NPS sources, to sort out the sources of the pollutants identified, and to demonstrate the effectiveness of BMPs in management systems. These results are crucial to provide resource management agencies with the best possible information.

Monitoring is needed to (1) document baseline conditions, (2) track progress and identify possible modifications or refocusing of efforts, and (3) document water-quality improvements. Demonstrating improvement is essential to justify the resources invested both to the landowners and, perhaps more importantly, to the tax-paying public demanding improved water quality and effective agency programs.

REFERENCES

- Betson, R.P. and J.B. Marius. 1969. Source areas of storm runoff.

 Water Resources Research 5(3):574-582.
- Byl, T.D. and A.J. Roman-Mas. 1994. Biomonitoring an agricultural watershed in West Tennessee. *in* Responses to Changing Multiple-use Demands: New Directions for Water Resources Planning and Management, AWRA Symposium, eds. M.J. Sale and R.O. Wadlington. pp 305-310.
- Cooper, C.M. 1993. Biological effects of agriculturally derived surface water pollutants on aquatic systems A review. J. Environ. Qual. 22:402-408.

- Dabney, S.M., K.C. McGregor, L.D. Maeyer, and G. Foster. 1993.

 Vegetative barriers for runoff and sediment control.

 Proceedings from the Tennessee Soil and Water Research and Education Committee, MTSU, Murfeesboro, TN.
- Griffin, M., W. Kreutzberger and P. Binney. 1992. Research needs for nonpoint source impacts. Water Environ. Tech. June, pp 60-64.
- Hamlett, J.M. and D.J. Epp. 1994. Water quality impacts of conservation and nutrient management practices in Pennsylvania. J. Soil Water Conserv. 49(1):59-66.
- Jayachandran, K., T.R. Steinheimer, L. Somasundaram, T.B. Moorman, R.S. Kanawar, and J.R. Coats. 1993. Distribution of atrazine and metabolites as contaminants in groundwater and soil cores from no-till corn. in Agricultural Research to Protect Water Quality: Proceedings of the Conference. Sponsored by the Soil and Water Conservation Society, Ankeny, IA, pp 105-109.
- Knopman, D.S. and R.A. Smith. 1993. Twenty years of the Clean
 Water Act. Environment 35(1):17-41.
- Richards, R.P. and D.B. Baker. 1993. Pesticide concentration patterns in agricultural drainage networks in the Lake Erie basin. *Environ. Toxicol. Chem.* 12:13-26.
- Sharpley, A.N. 1981. The contribution of phosphorous leached from crop canopy to losses in surface runoff. *J. Environ. Qual.* 10(2):160-165.
- Sharpley, A.N., S.J. Smith and J.R. Williams. 1988. Nonpoint source pollution impacts of agricultural land use. Lake Reservoir Management 4(1):41-49.
- Sharpley, A.N., S.J. Smith, J.R. Williams, O.R. Jones and G.A. Coleman. 1991. Water quality impacts associated with sorghum culture in the southern plains. *J. Environ. Qual.* 20:239-244.
- Soileau, J.M., J.T. Touchton, B.F. Hajek, and K.H. Yoo. 1994.

 Sediment, nitrogen, phosphorous runoff with conventional- and conservation-tillage cotton in a small watershed. *J. Soil Water Conserv.* 49(1):82-89.
- Thompson, J. 1993. Non-point source pollution It points to all of us. The Tennessee Conservationist. vol. LIX, (5):6-9.